



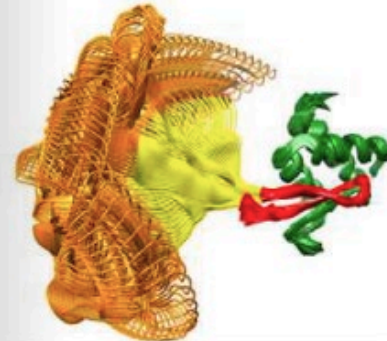
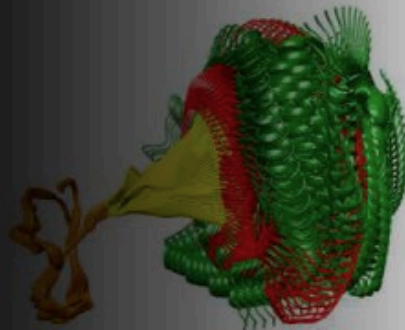
# The Impact of Deep Learning for Creating Novel Effectors of Biological Functions

**RPI STRUCTURAL  
BIOINFORMATICS  
LABORATORY**

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5th National Big Data Health  
Science Conference  
Columbia, South Carolina

February 3, 2024

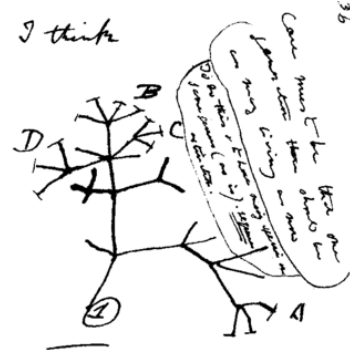
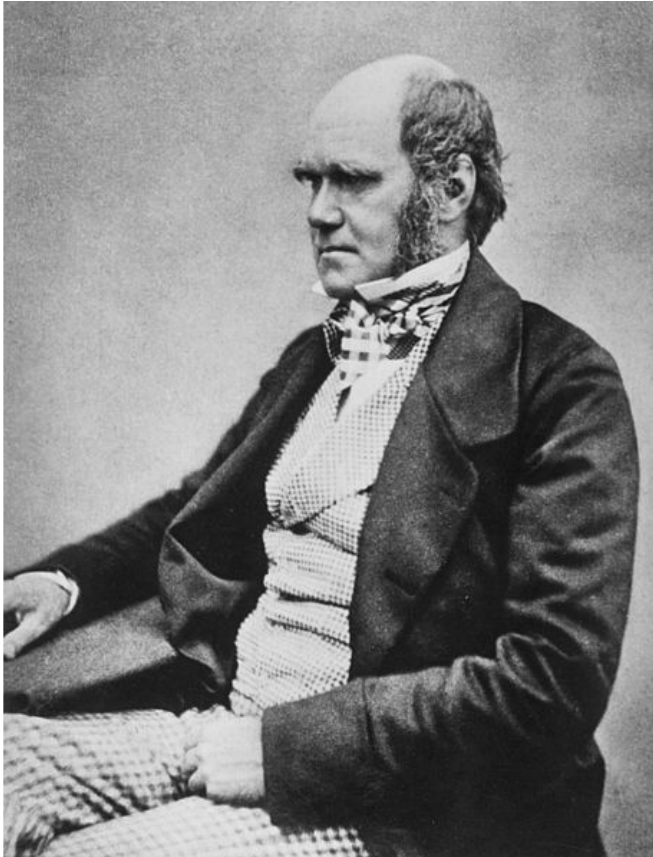


# BIG DATA

# Charles Darwin

## (1809-1882)

### Transmutation of Species



Then between A & B. various  
 sort of relation. C & B. the  
 first gradation, B & D  
 rather greater distinction  
 than former would be  
 formed. - binary relation

“Survival of the Fittest”

# Charles Darwin – Natural Selection

The concept of fitness is central to natural selection. In broad terms, individuals that are more "fit" have better potential for survival, as in the well-known phrase "survival of the fittest".

Proposed in parallel by Alfred Russell Wallace

The concept of natural selection was originally developed in the absence of a valid theory of heredity; at the time of Darwin's writing, nothing was known of modern genetics.



# Gregor Mendel

## (1822 – 1884)



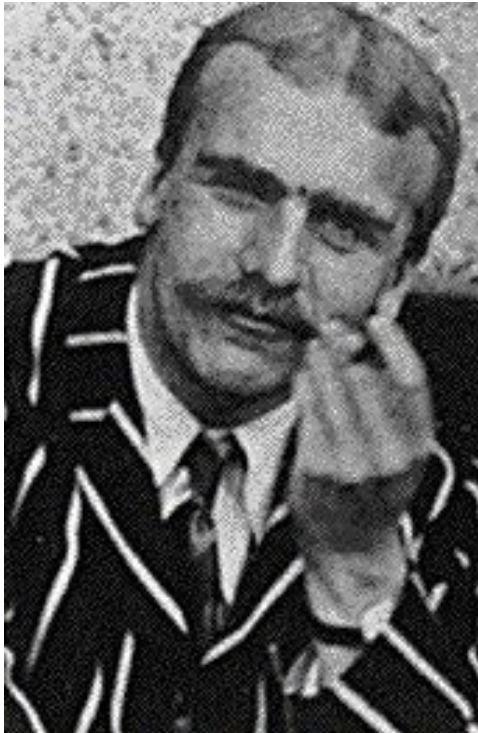
Mendel demonstrated that the inheritance of certain traits in pea plants follows particular patterns, now referred to as the laws of Mendelian inheritance.

“Particulate” nature of inheritance of traits.

The laws formed the foundation of the modern science of genetics

# J.B.S. Haldane

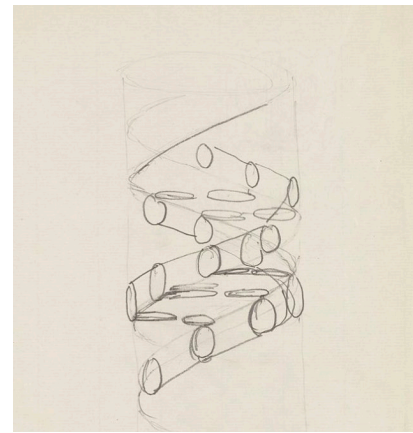
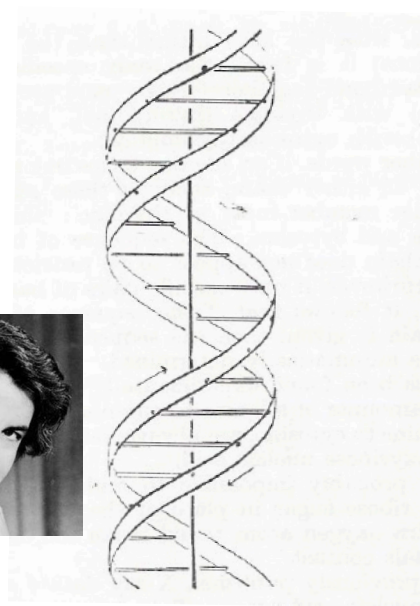
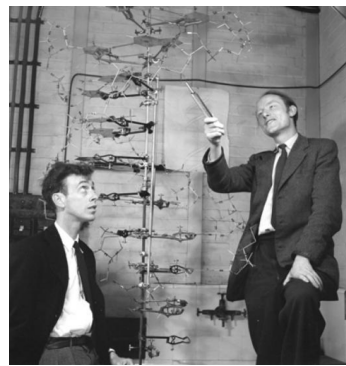
(1892– 1964)



British-born geneticist, protein chemist, and evolutionary biologist generally credited with a central role in the development of **neo-Darwinian** thinking .

Genetic (gene) – basis of transmutation of species

Mutations of genes give rise to “transmutation of species” by changing the structure of the encoded gene product

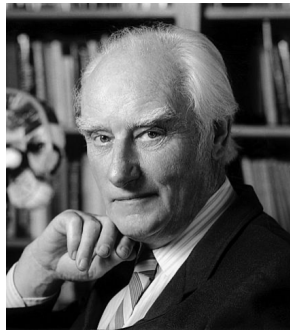


**In February 1953, Watson and Crick completed their model, which is now accepted as the first correct model of the double-helix of DNA.**

**On 28 February 1953 Crick interrupted patrons' lunchtime at The Eagle Pub in Cambridge to announce that he and Watson had "discovered the secret of life"**

One strand templates  
the second strand

Francis Crick  
James Watson  
Rosalind Franklin



Francis Crick in 1957

**“Structural Biology” – using  
molecular structures to  
understand biology**

# Paradigm of Molecular Biology

DNA Sequence



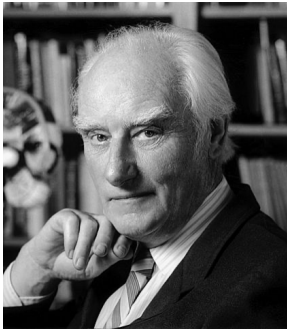
transcription

RNA Sequence



translation

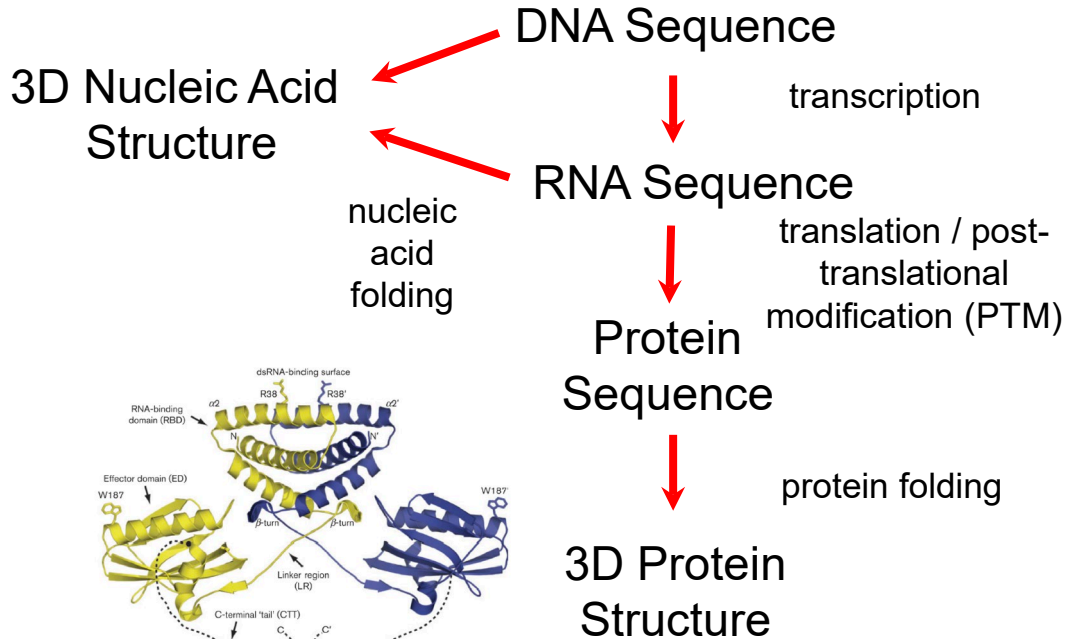
Protein  
Sequence



Francis Crick in 1957

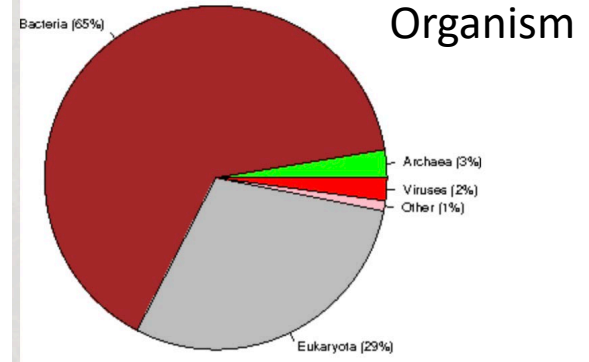
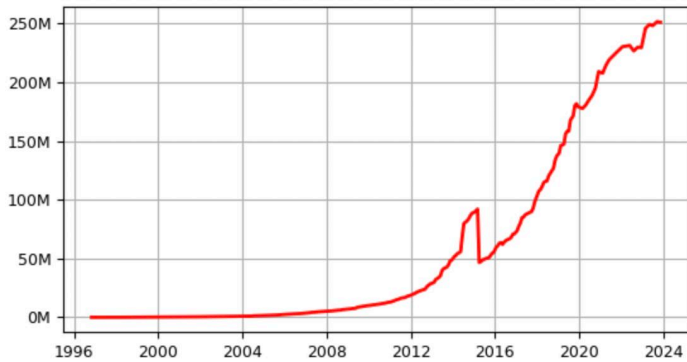
In 1979, Alexander Rich and co-workers at MIT grew a crystal of Z-DNA. This was the first crystal structure of any form of DNA.

# Paradigm of Molecular Biology

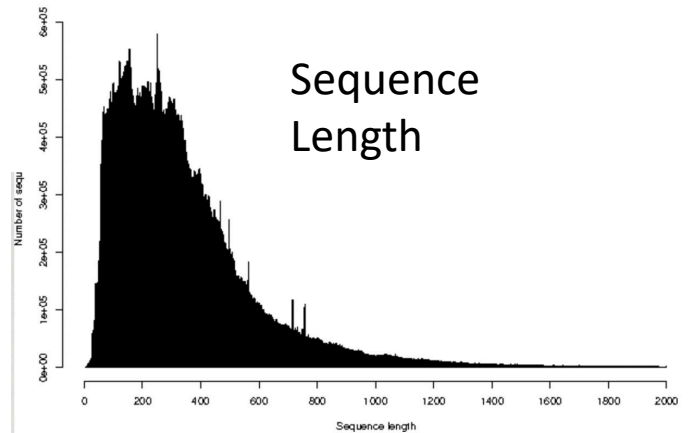


X-ray crystallography  
Nuclear Magnetic Resonance (NMR)  
Cryo Electron Microscopy

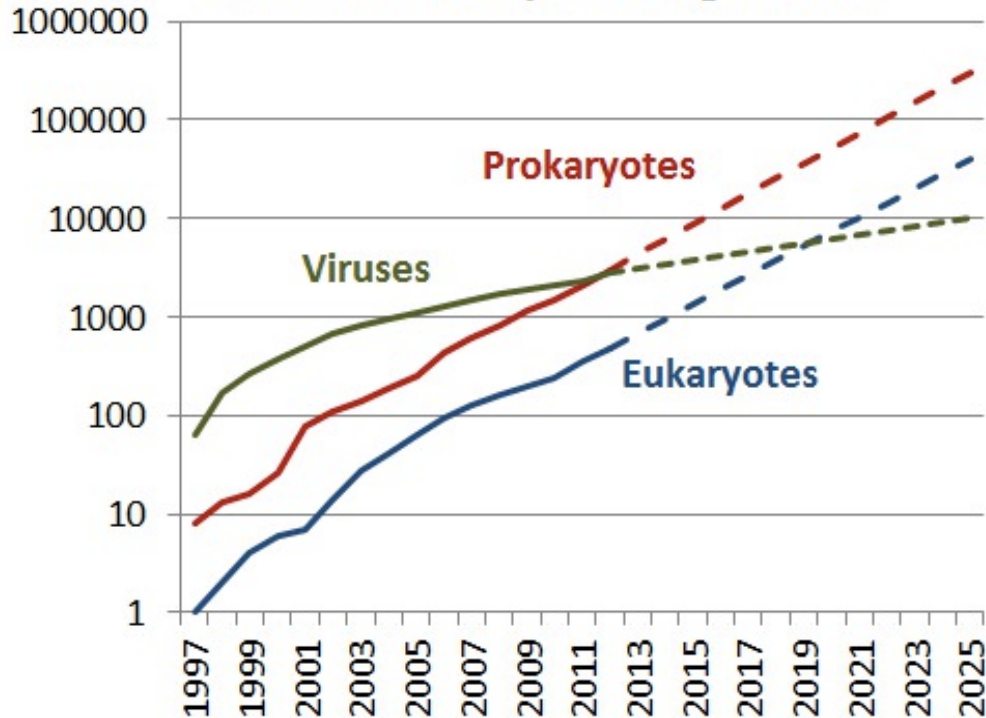
Number of entries in UniProtKB/TrEMBL



Length distribution of the sequences



## Cumulative sequenced genomes



The number of genomes sequenced increases by > 10-fold every 5 years

Over the next 15 yrs we expect to have 1,000 -fold more sequences than we have today.

~~1 Gene → 1 Protein~~

**1 Gene → Multiple Proteins**

- Splicing
- Proteolytic Processing
- Posttranslational modifications
  - glycosylation
  - phosphorylation
  - acetylation
  - methylation
  - ubiquitination and sumylation
  - other modifications



# Protein Structure Databank (PDB)

RCSB **PDB** PROTEIN DATA BANK

214,226 Structures from the PDB  
1,068,577 Computed Structure Models (CSM)

3D Structures  Include CSM

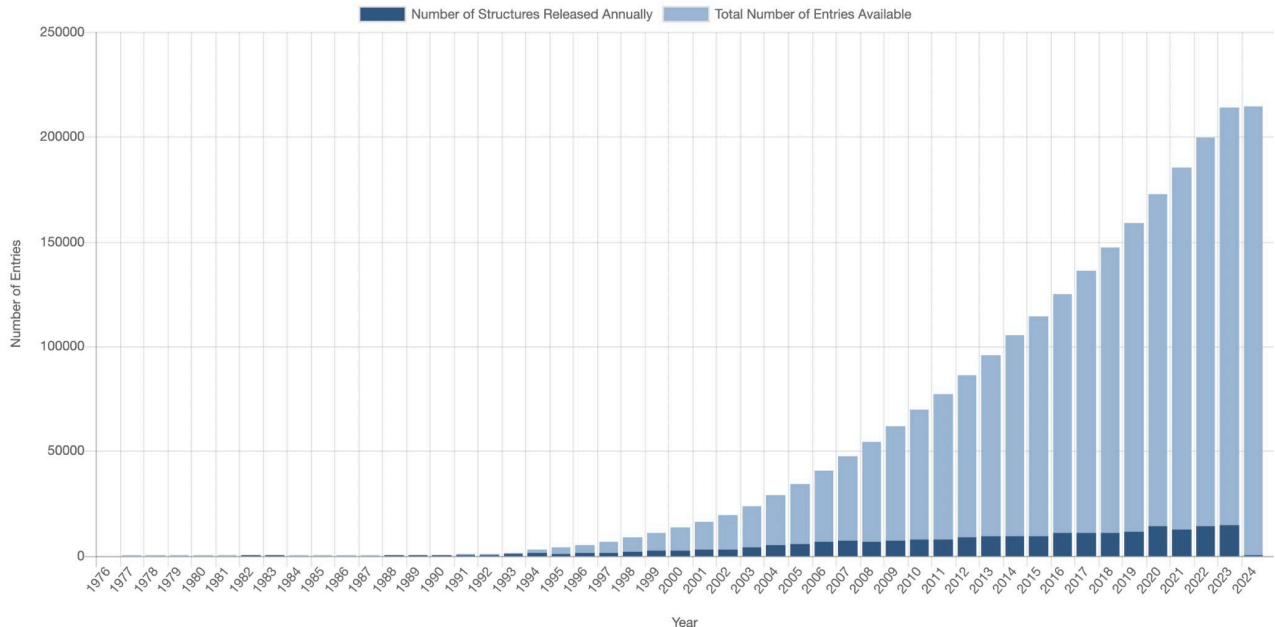
[Advanced Search](#) | [Browse Annotations](#) [Help](#)

[PDB-101](#) [PDB](#) [EMDataResource](#) [NAKB](#) [wwPDB Foundation](#) [PDB-Dev](#)

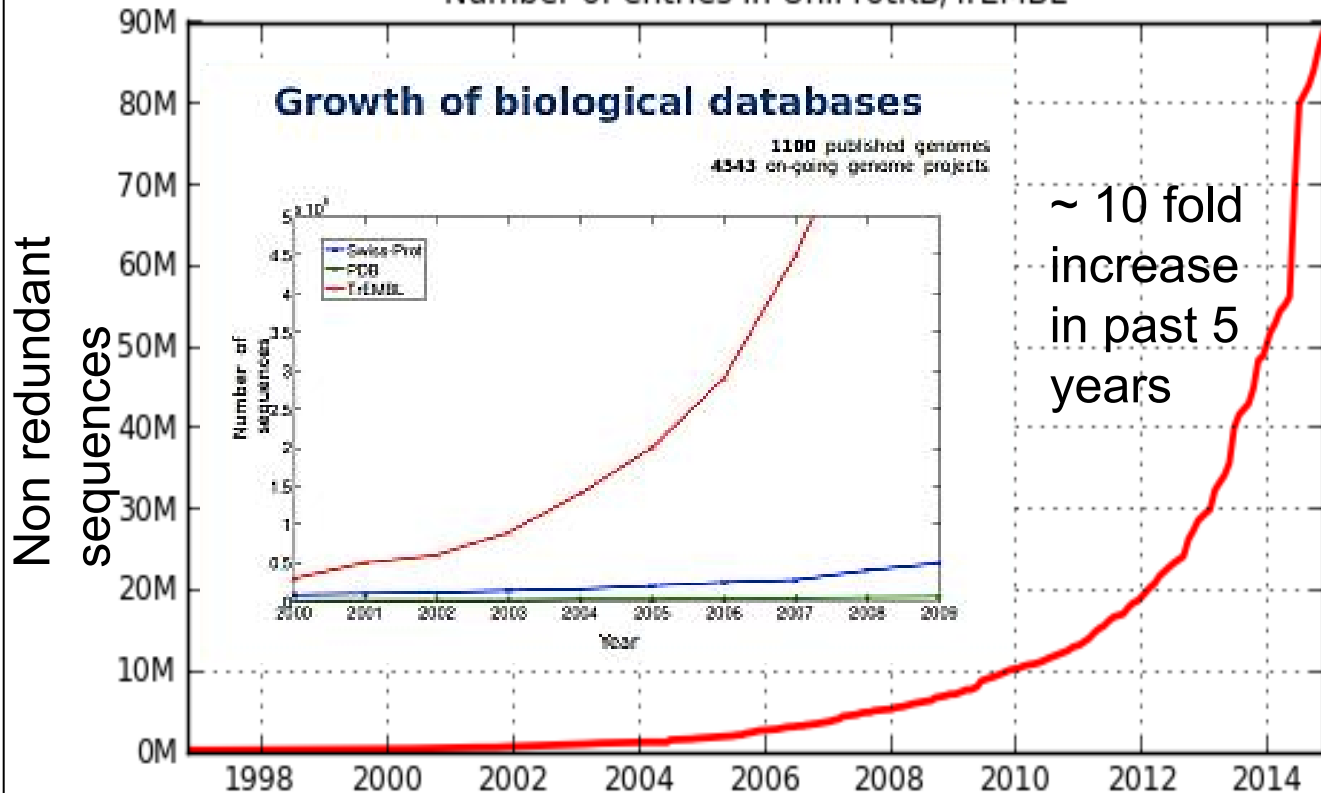
[f](#) [t](#) [v](#) [y](#)

## PDB Statistics: Overall Growth of Released Structures Per Year

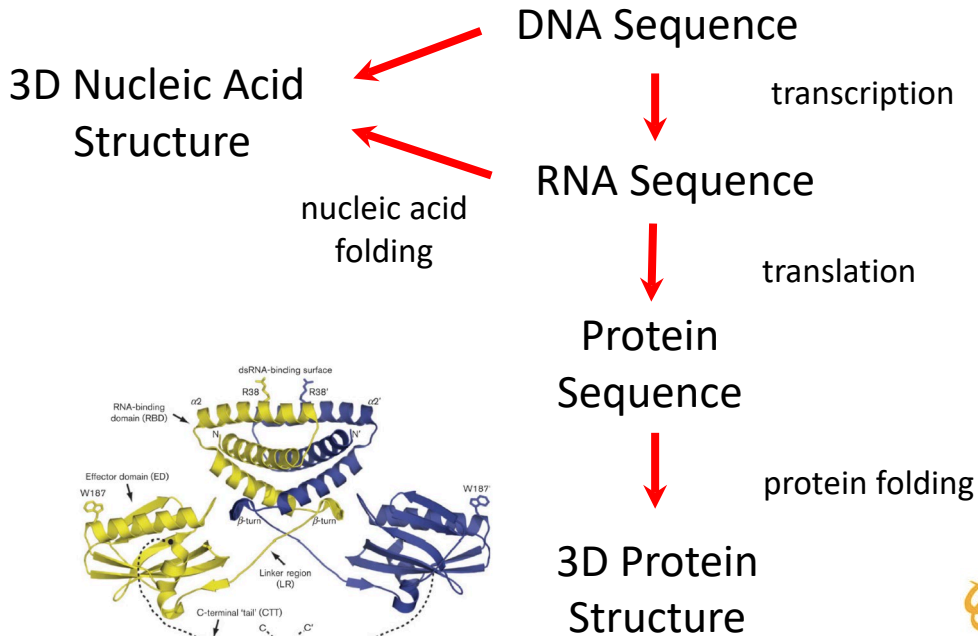
All Statistics



# Number of entries in UniProtKB/TrEMBL



# Paradigm of Molecular Biology



## International Protein Structure Initiative

The long-range goal of the Protein Structure Initiative (PSI) is to make the three-dimensional atomic-level structures of most proteins easily obtainable from knowledge of their corresponding DNA sequences.

# Structural genomics and the Protein Data Bank

Received for publication, February 17, 2021, and in revised form, April 16, 2021 Published, Papers in Press, May 3, 2021,  
<https://doi.org/10.1016/j.jbc.2021.100747>

Karolina Michalska<sup>1,2</sup> and Andrzej Joachimiak<sup>1,2,3,\*</sup> 

By the end of the PSI program (2000 – 2016), more than 9400 structures determined, with the majority of them being unique.

**Table 1**  
**Top 20 structural genomics programs**

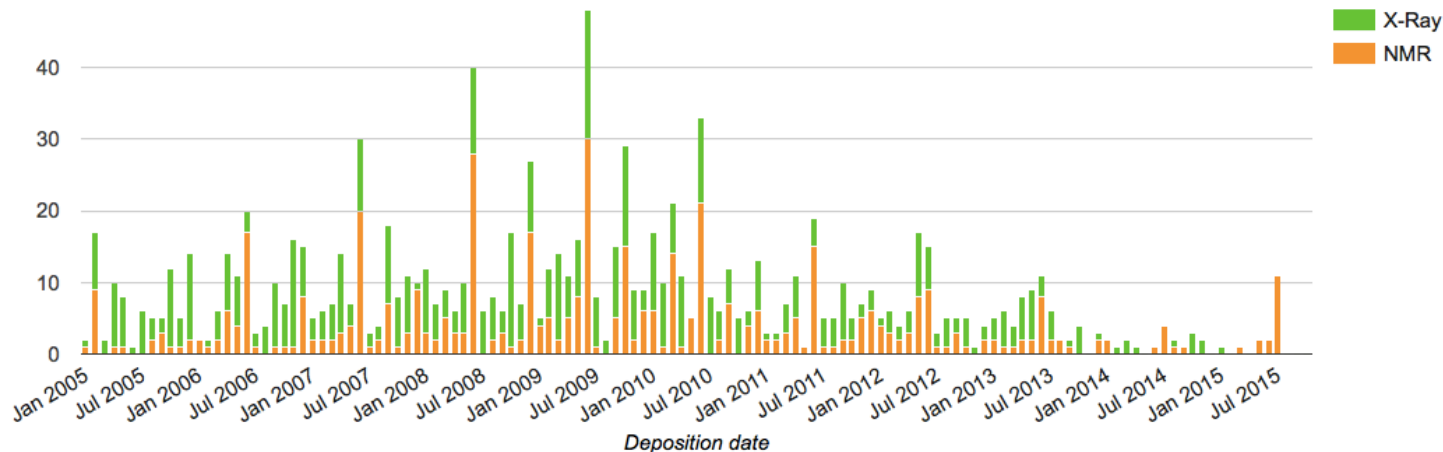
Center	Number of PDB deposits	Origin and funding	Techniques used
RIKEN Structural Genomics/Proteomics Initiative	2746	Japan, government, National Project on Protein Structural and Functional Analyses	NMR, X-ray
Midwest Center for Structural Genomics	1955	USA, PSI/NIH/NIGMS	X-ray, NMR
Structural Genomics Consortium	1896	International/a public-private partnership	X-ray, NMR
Joint Center for Structural Genomics	1601	USA, PSI/NIH/NIGMS	X-ray, NMR
Center for Structural Genomics of Infectious Diseases	1359	USA, NIH/NIAID	X-ray, NMR, cryo-EM
Seattle Structural Genomics Center for Infectious Disease	1355	USA, NIH/NIAID	X-ray, NMR, cryo-EM
Northeast Structural Genomics Consortium	1234	USA, PSI/NIH/NIGMS	X-ray, NMR
New York SGX Research Center for Structural Genomics	1041	USA, PSI/NIH/NIGMS	X-ray, NMR
New York Structural Genomics Research Consortium	364	USA, PSI/NIH/NIGMS	X-ray, NMR
TB Structural Genomics Consortium	344	International worldwide consortium/Various	X-ray, NMR
Center for Eukaryotic Structural Genomics	219	USA, PSI/NIH/NIGMS	X-ray, NMR
Montreal-Kingston Bacterial Structural Genomics Initiative	132	Canada, Canadian Institutes of Health Research	X-ray, NMR
Southeast Collaboratory for Structural Genomics	122	USA, PSI/NIH/NIGMS	X-ray, NMR
Structural Proteomics in Europe	118	European Union	X-ray, NMR
Berkeley Structural Genomics Center	101	USA, PSI/NIH/NIGMS	X-ray
Enzyme Discovery for Natural Product Biosynthesis	91	USA, NIH	X-ray
Structural Genomics of Pathogenic Protozoa Consortium	73	USA, PSI/NIH/NIGMS	X-ray, NMR
New York Consortium on Membrane Protein Structure	70	USA, PSI/NIH/NIGMS	X-ray
Structure 2 Function Project	54	USA, PSI/NIH/NIGMS	X-ray, NMR
GPCR Network	52	USA, PSI/NIH/NIGMS	X-ray

NIAID, National Institute of Allergy and Infectious Diseases; NIGMS, National Institute of General Medical Sciences; NIH, National Institutes of Health; PSI, Protein Structure Initiative.



NMR: 570 structures  
X-Ray: 664 structures  
Total: 1,234 structures  
177 Euka; 150 Human  
> 200,000 homology models

NESG Monthly Structure Depositions

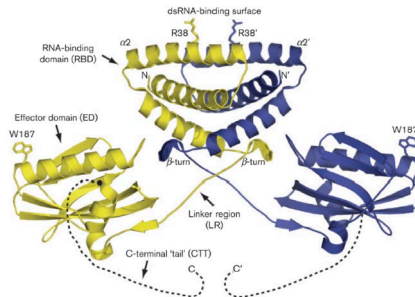


**NMR:** Arrowsmith, Kennedy, Montelione Powers, Prestegard, Szyperski, Valafar (Aramini, Cort, Eletsy, Gutmanas, Lamack, Lee, Liu, Mani, Mercier, Mills, Pederson, Pulavarti, Ramelot, Rossi, Singarapu, Shen, Stark, Swapna, Tang, Wu, Xu, Yang, Yee and others)

**X-ray:** Hunt, Tong, Montelione labs

# Paradigm of Molecular Biology

Can we do this ALL  
*in silico*



DNA Sequence



transcription

RNA Sequence



translation

Protein  
Sequence



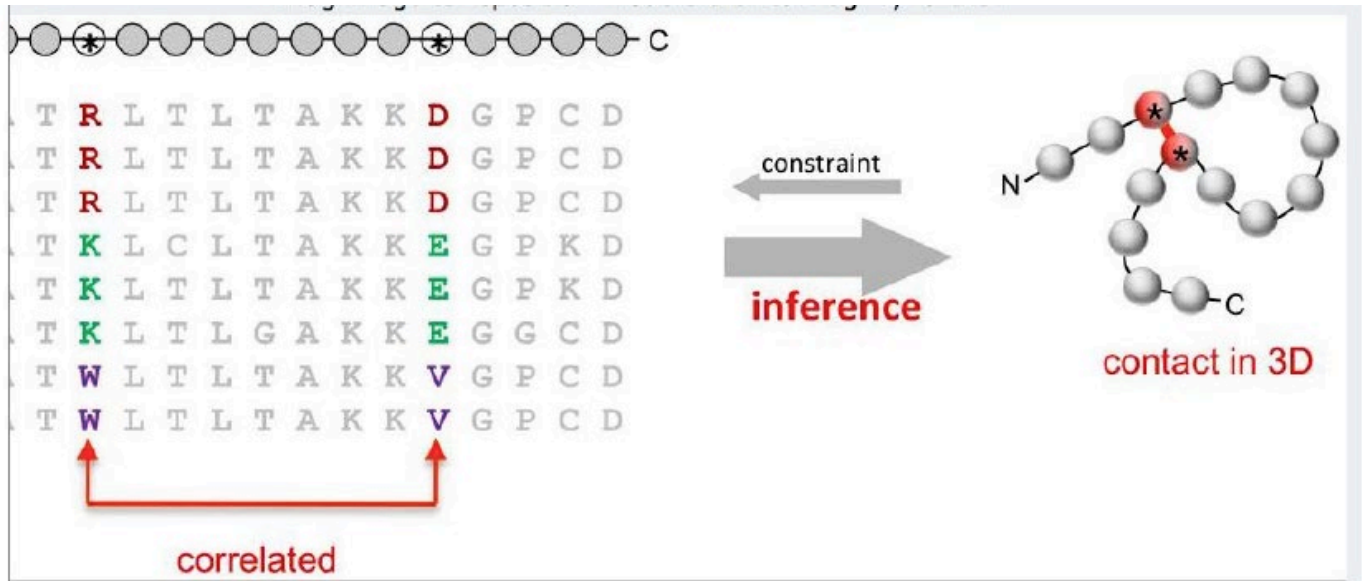
protein folding /  
PTM

3D Protein  
Structure



Funded by the National  
Institutes of Health

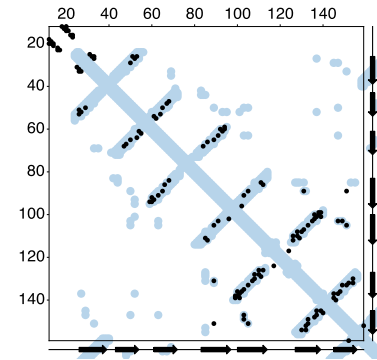
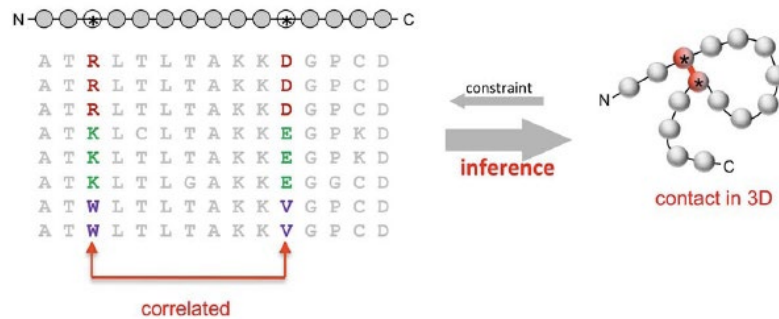
# Evolutionary Co-Variance (EC)



# Direct-coupling analysis of residue coevolution captures native contacts across many protein families

Faruck Morcos<sup>a,1</sup>, Andrea Pagnani<sup>b,1</sup>, Bryan Lunt<sup>a</sup>, Arianna Bertolino<sup>c</sup>, Debora S. Marks<sup>d</sup>, Chris Sander<sup>e</sup>,  
Riccardo Zecchina<sup>b,f</sup>, José N. Onuchic<sup>a,g,2</sup>, Terence Hwa<sup>a,2</sup>, and Martin Weigt<sup>b,h,2</sup>

PNAS, 110, 20533 2011





# Protein 3D Structure Computed from Evolutionary Sequence Variation

Debora S. Marks<sup>1,2,3</sup>, Lucy J. Colwell<sup>2,3</sup>, Robert Sheridan<sup>3</sup>, Thomas A. Hopf<sup>1</sup>, Andrea Pagnani<sup>4</sup>, Riccardo Zecchina<sup>4,5</sup>, Chris Sander<sup>3</sup>

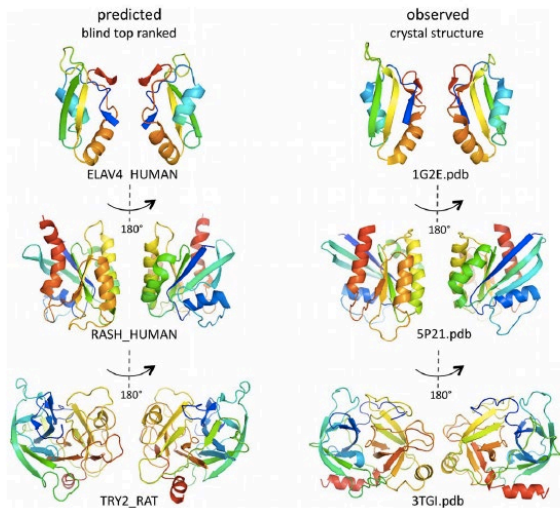


Table 1. Accuracy of predicted proteins.

Target Protein Uniprot ID	Fold	L*	Pfam ID	No. seqs	Blind top Ca-rmsd**	TM***	Best Ca-rmsd**	TM***	TP****	Ref. PDB
RASH_HUMAN	a/b	161	Ras	10K	3.5 (161)	0.7	2.8 (155)	0.76	0.8	5p21
CHEY_ECOLI	a/b	114	Response_reg	72K	2.98 (107)	0.65	2.96 (107)	0.67	0.67	1e6k
THIO_ALIAC	a/b	103	Thioredoxin	13K	3.86 (94)	0.55	3.5 (97)	0.59	0.68	1rqm
RNH_ECOLI	a/b	141	RNase_H	11K	4.0 (110)	0.54	3.5 (114)	0.57	0.68	1f21
TRY2_RAT	b	223	Trypsin	16K	4.27 (186)	0.6	4.27 (186)	0.54	0.81	3tgi
CADH1_HUMAN	b	100	Cadherin	12K	3.8 (88)	0.55	3.86 (96)	0.57	0.86	2o72
YES_HUMAN	b	48	SHB_1	6K	3.6 (47)	0.37	3.35 (43)	0.41	0.52	2hda
O45418_CAEEL	a+b	100	FKBP_C	8K	4.1 (88)	0.48	3.4 (79)	0.53	0.77	1r9h
ELAV4_HUMAN	a+b	71	RRM_1	28K	2.9 (67)	0.57	3.16 (71)	0.59	0.71	1g2e
A8MVQ9_HUMAN	a+b	107	Lectin_C	5K	4.8 (85)	0.39	4.0 (100)	0.53	0.8	2n6
PCBP1_HUMAN	a+b	63	KH_1	9K	4.69 (46)	0.25	4.61 (61)	0.35	0.47	1wvn
OPSD_BOVIN	a tm	258	7tm_1	27K	4.84 (171)	0.5	4.29 (180)	0.55	0.38	1hcx
BPT1_BOVIN	a+b	52	Kunitz_BPT1	2K	2.73 (53)	0.49	2.75 (53)	0.49	0.71	5p2i
OMPR_ECOLI	a	77	Trans_reg_C	24K	4.7 (64)	0.35	3.9 (62)	0.45	0.38	1odd
SPTB2_HUMAN	a	108	CH(calp hom)	4K	4.0 (47)	0.37	3.88 (88)	0.5	0.5	1blr

Backbone RMSD of top model to Xtal structure of 3 – 5 Å

# Genomics-aided structure prediction

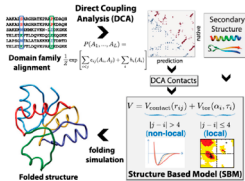
Joanna I. Sułkowska<sup>a,1</sup>, Faruck Morcos<sup>a,1,2</sup>, Martin Weigt<sup>b</sup>, Terence Hwa<sup>a,2</sup>, and José N. Onuchic<sup>c,2</sup>

<sup>a</sup>Center for Theoretical Biological Physics, University of California at San Diego, La Jolla, CA 92093-0374; <sup>b</sup>Laboratoire de Génie Microorganismes, UMR 7238, Université Pierre et Marie Curie, 15 rue de l'École de Médecine, 75006 Paris, France; and <sup>c</sup>Center for Theoretical Biological Physics, Rice University, Houston, TX 77005-1827

Contributed by José N. Onuchic, May 9, 2012 (sent for review January 20, 2012)

DCA-fold, integrating DCA contacts with an accurate knowledge of local information is sufficient to fold proteins in the range of 1–3 Å accuracy

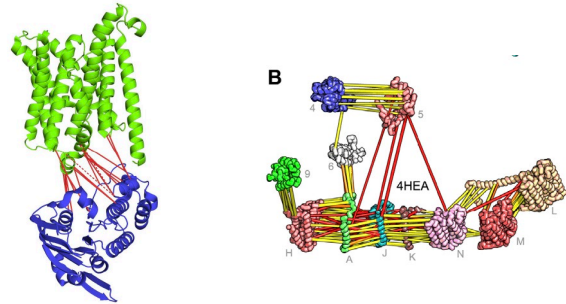
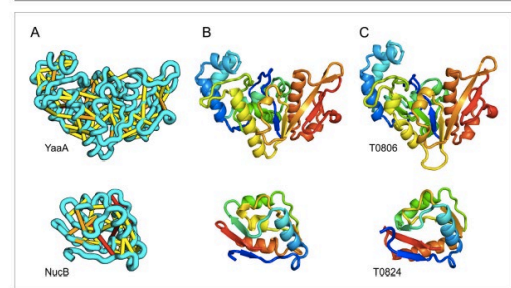
PNAS 2012



# Large-scale determination of previously unsolved protein structures using evolutionary information

Sergey Ovchinnikov<sup>1</sup>, Lisa Kinch<sup>2</sup>, Hahnbeom Park<sup>1</sup>, Yuxing Liao<sup>3</sup>, Jimin Pei<sup>2</sup>, David E Kim<sup>1</sup>, Hetunandan Kamisetty<sup>4</sup>, Nick V Grishin<sup>2,3</sup>, David Baker<sup>1,5\*</sup>

EC restrained Rosetta: Backbone RMSD of top models to Xtal structure of 2.7 – 6 Å  
eLife 2015; Science 2017



BtuC – BtuD



RESEARCH ARTICLE



2014

## Sequence co-evolution gives 3D contacts and structures of protein complexes

Thomas A Hopf<sup>1,2†</sup>, Charlotta P I Schärfe<sup>1,2,4†</sup>, João P G L M Rodrigues<sup>5†</sup>, Anna G Green<sup>1</sup>, Oliver Kohlbacher<sup>3,4</sup>, Chris Sander<sup>6\*</sup>, Alexandre M J J Bonvin<sup>5\*</sup>, Debora S Marks<sup>1\*</sup>



RESEARCH ARTICLE



2014

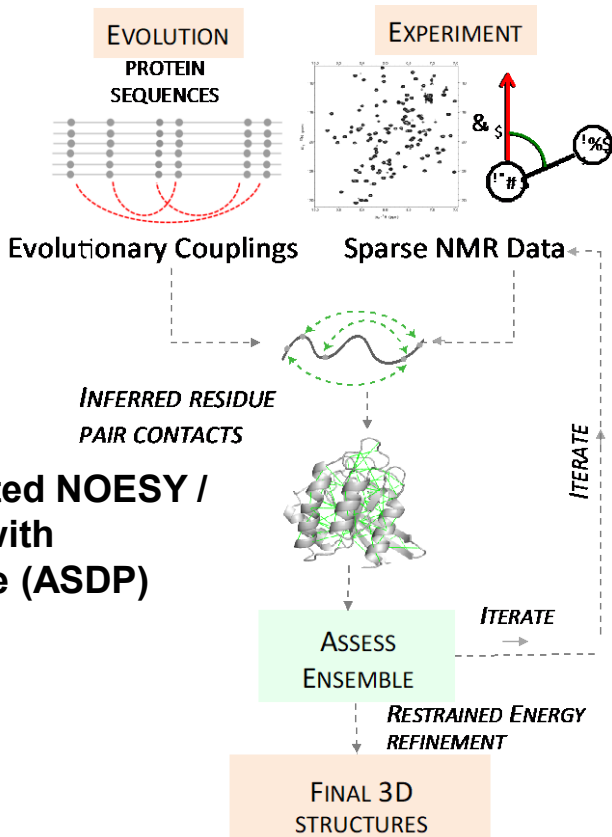
## Robust and accurate prediction of residue–residue interactions across protein interfaces using evolutionary information

Sergey Ovchinnikov<sup>1,2†</sup>, Hetunandan Kamisetty<sup>1,3†</sup>, David Baker<sup>1\*</sup>

# Can we enable protein structure determination by combining sparse NMR data with EC restraints?

Yuefeng Tang  
Janet Huang  
Thomas Hopf  
Chris Sander  
Debora Marks

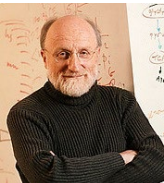
## EC-NMR Method



Structure generation

Refined Residue Pair Contacts -> Atom Pair Contacts

Restrained Rosetta

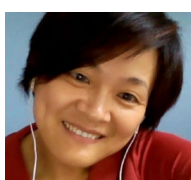


C Sander

D Marks

EVFold Server

Fully automated NOESY / EC analysis with AutoStructure (ASDP)



Tang, Huang et al  
Nature Methods 2015

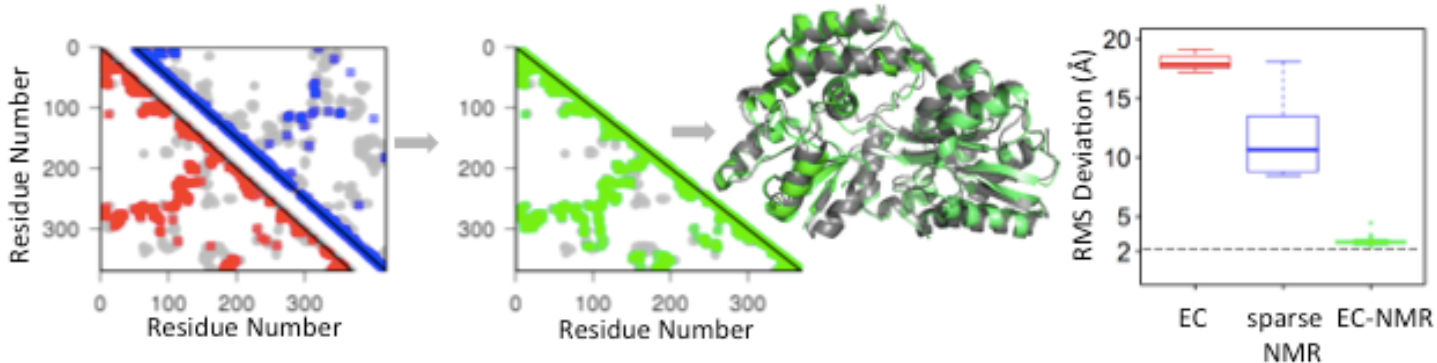
# Hybrid EC-NMR Method

$H^N$ , ILV Me Assignments

1  $^{15}N$ - $^1H$  RDC alignment

$N_{\text{eff}} = 12,416$        $N_{\text{eff}} / L = 33$

Maltose Binding Protein 370 residues 41.9 kD



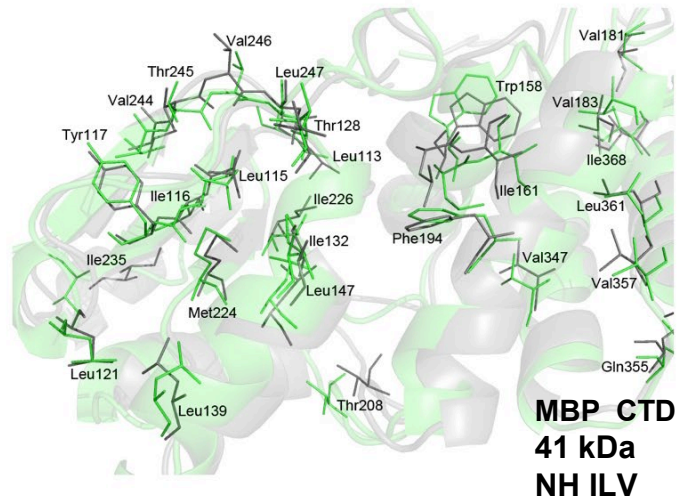
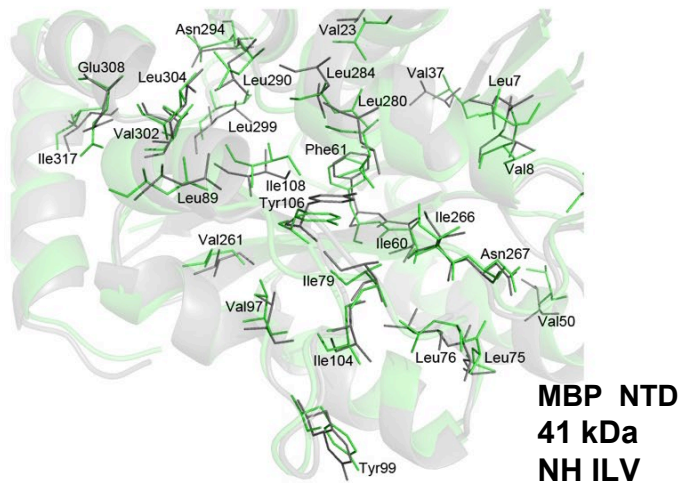
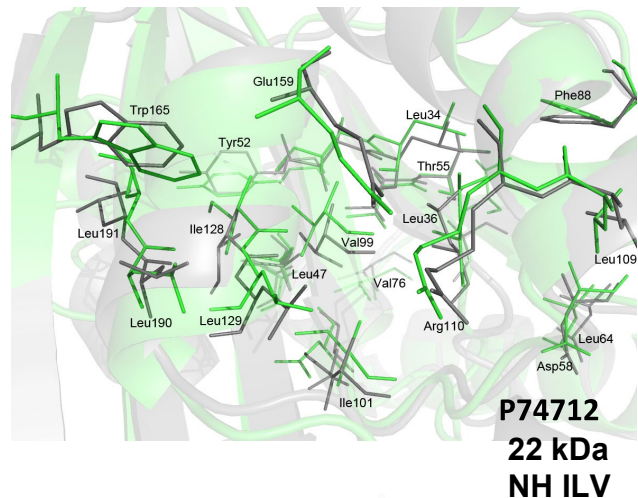
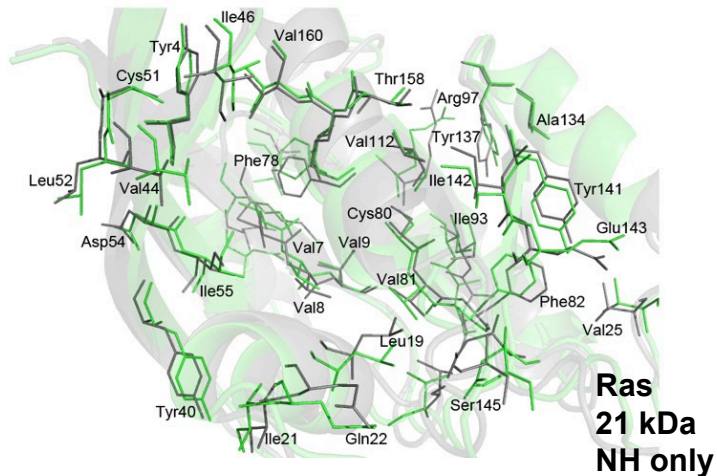
NOESY cross peak assignment ambiguity is iteratively resolved

False positive ECs are identified and removed

ECs are deconvolved into atom-atom distance restraints.

MBP – beta cyclodextrin complexes

# EC-NMR structures have accurate core sidechain structures





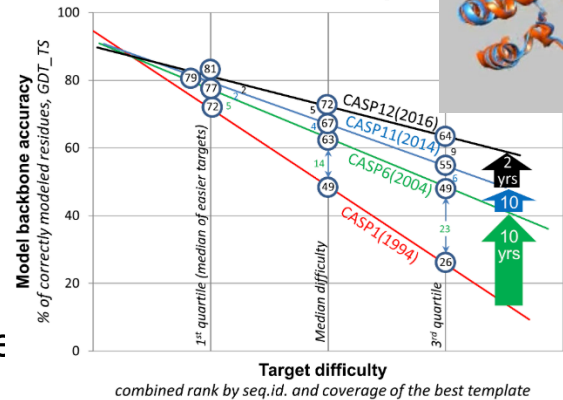
# CASP – Critical Assessment of Protein Structure Prediction

A community-wide, worldwide experiment for protein structure prediction taking place every two years since 1994

CASP provides research groups with an opportunity to objectively test their structure prediction methods and delivers independent assessment of the state of the art in protein structure modeling

More than 100 research groups from all over the world participate in CASP on a regular basis

<http://predictioncenter.org/>



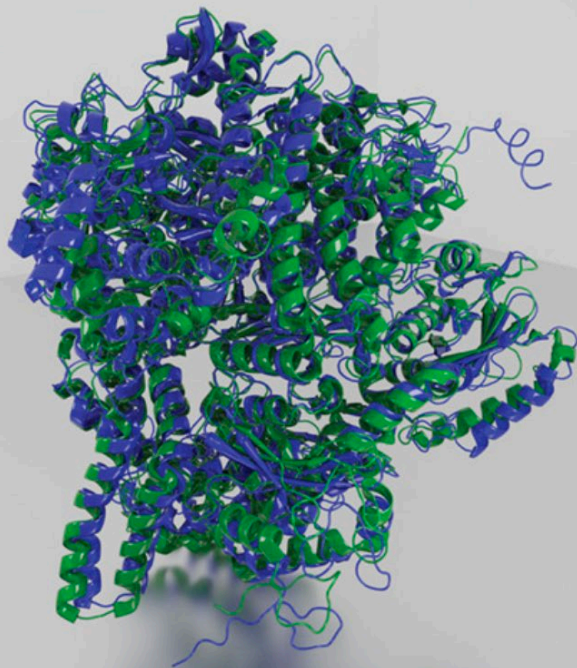
Categories:

- Template-based Modeling
- ab initio modeling
- contact prediction
- Refinement
- Data-assisted prediction

# PROTEINS

STRUCTURE ■ FUNCTION ■ BIOINFORMATICS

Special issue: CASP14: Critical Assessment of methods of protein Structure Prediction, 14th round.  
Edited by John Moult and Andriy Kryshtafovych



WILEY

## CASP – Critical Assessment of Protein Structure Prediction

- Experimental Structures
- Sequences Distributed
- Blind Predictions
- Third-party Assessors

CASP1 – 1994

...

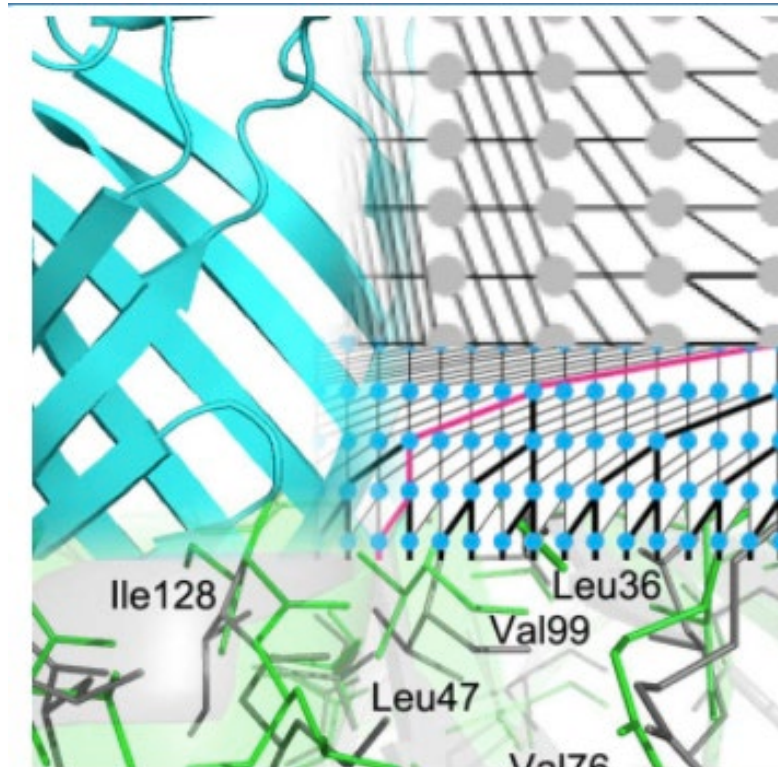
CASP15 - 2022

**rmsd** – root-mean square deviation of atomic coordinates.

2 crystal structures < 1.5 Å

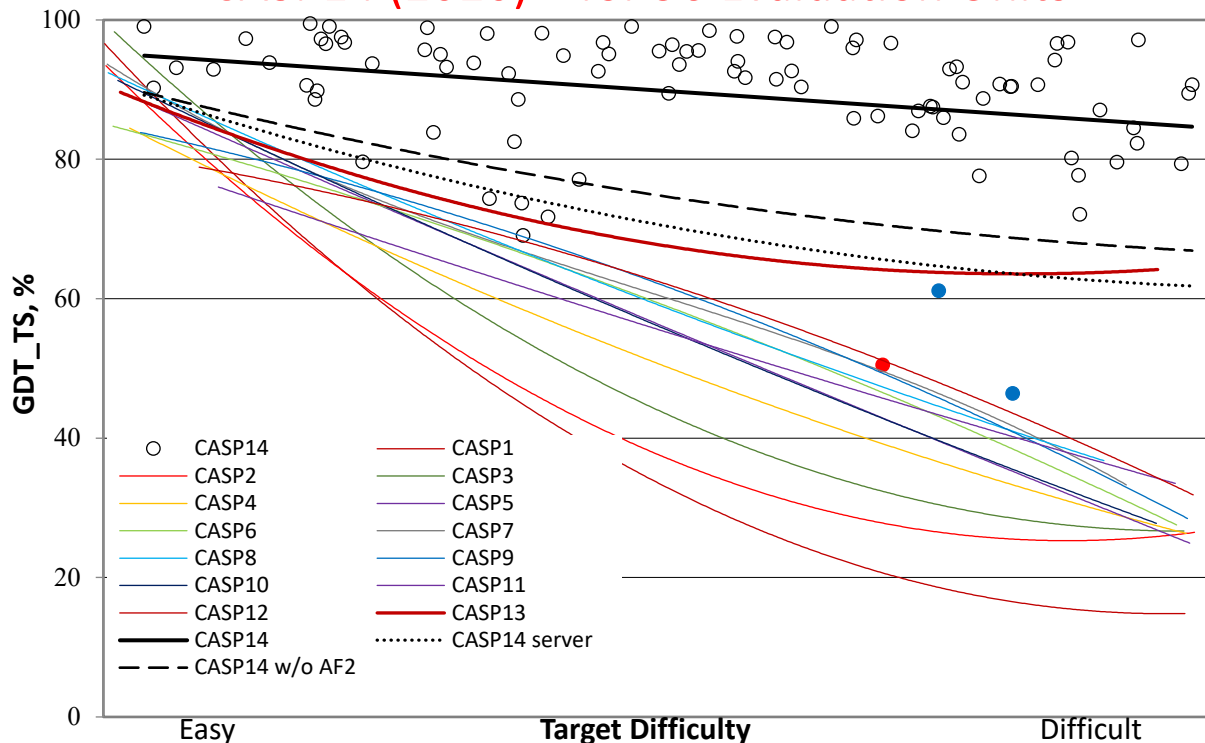
**GDT** – global distance test. Percent of atoms < 1 Å rmsd

# Deep Learning





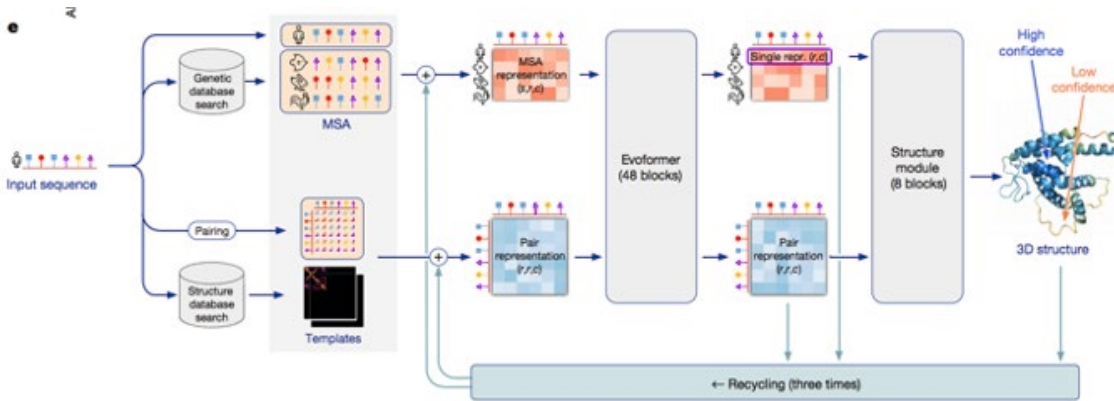
# Remarkable Performance of AlphaFold in CASP14 (2020) – for 96 Evaluation Units



For 96 CASP14 targets AF2 models had a mean GDT\_TS of  $0.88 \pm 0.1$ , corresponding to RMSD between predicted and experimental protein structures of about 1.5 Å

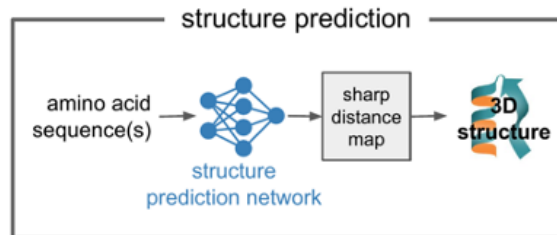
Kryshtafovych, Fidelis, Moulton et al, 2021

# Highly Accurate Protein Structure Prediction with AlphaFold: Deep Learning from the Protein Structure DataBase



Jumper et al Nature 2021

Deep Mind,  
Inc

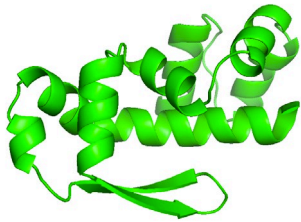


AlphaFold –  
Convolutional  
Neural Network

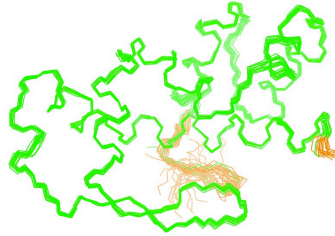
AlphaFold2 –  
Attention-based  
Network

# AlphaFold 2020

## CASP14 Target T1055



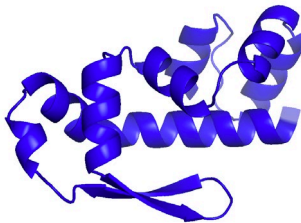
NMR  
Model



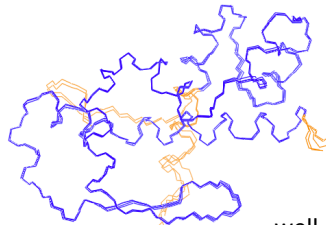
NMR Ensemble

well-defined region:  
residues 305 - 426

Common Region for RMSD /  
GDT comparison: 310-426



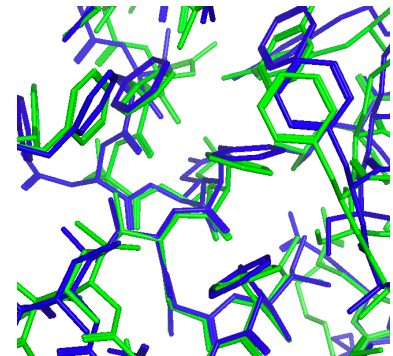
AF Model



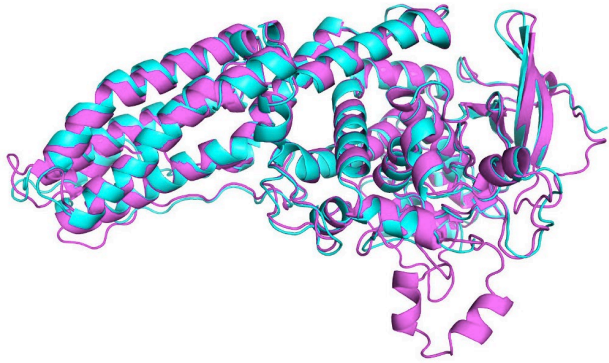
AF Ensemble

well-defined region:  
residues 310-428

$\langle \text{RMSD} \rangle = 0.97 \text{ \AA}$   
GDT = 0.90



Huang YJ, Zhang N, Bersch B, Fidelis K, Inouye M, Ishida Y, Kryshstafovych A, Kobayashi N, Kuroda Y, Liu G, LiWang A, Swapna GVT, Wu N, Yamazaki T, Montelione GT. Assessment of prediction methods for protein structures determined by NMR in CASP14: Impact of AlphaFold2. **Proteins**. 2021



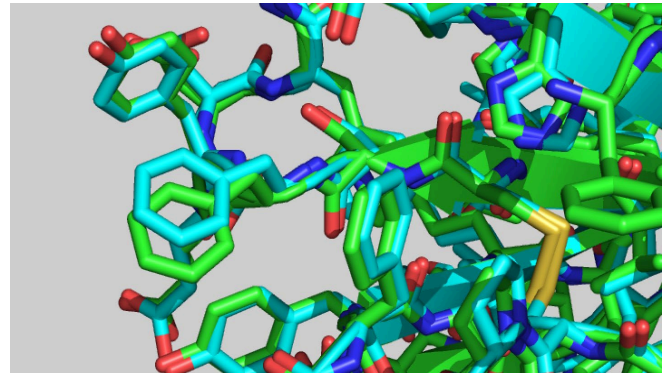
CASP target T1053, a two-domain bacterial kinase. Both domains are difficult modeling targets (FM/TBM category).

attention-based  
machine learning

largest CASP14 target  
was 949 res

Kryshtafovych et al, 2021

AF2 and RosettaFold  
source freely available



Dec 2020 CASP14

Jul 2021 AlphaFold (AF2) Published in *Nature*

Jul 2021 RoseTTAFold Published in *Science*

Aug 2021 AF and RoseTTAFold Servers *Google Cloud*

Aug 2021 20 Proteomes published in *Science*

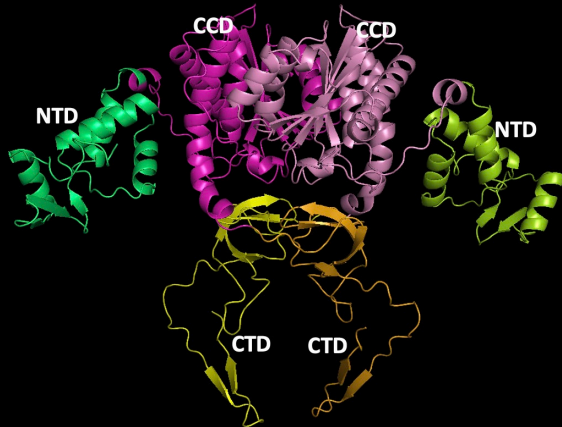
Oct 2021 AF Running at RPI on CCI GPU Clusters

Jul 2022 200 M structures released by Deep Mind

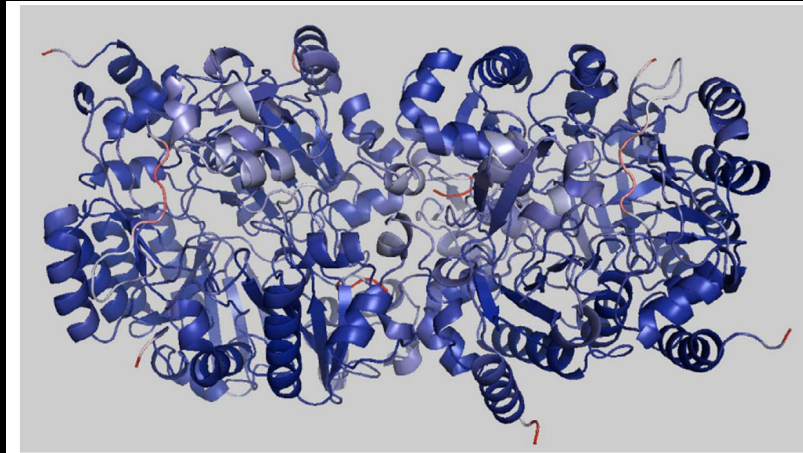
## MLV Integrase

408 aa X 2 = 816 aa ~ 90 kDa

Chain A in cyan and chain B in red



## Eury DsrAB Disulfide Reductase 1400 residues ~ 155 kDa



**FIGURE 4** AlphaFold2 model of MV2.Eury DsrAB. The sequence is colored for pLDDT score (described in the text), with dark blue corresponding to high confidence prediction (96%) and red to low confidence prediction (43%). The view is a top view of the structure

GVT Swapna  
C Royer

“We’re releasing now the structures for the whole protein universe,” said Demis Hassabis, founder and CEO of DeepMind, at a press conference in London.

## Science Magazine “Breakthrough of 2021”

Science

Current Issue First re

HOME > SCIENCE > VOL. 374, NO. 6574 > PROTEINS, PROTEINS EVERYWHERE

EDITORIAL

### Proteins, proteins everywhere

H. HOLDEN THORP

SCIENCE • 16 Dec 2021 • Vol 374, Issue 6574 • p. 1415 • DOI:10.1126/science.abn5795

NEWS | TECHNOLOGY

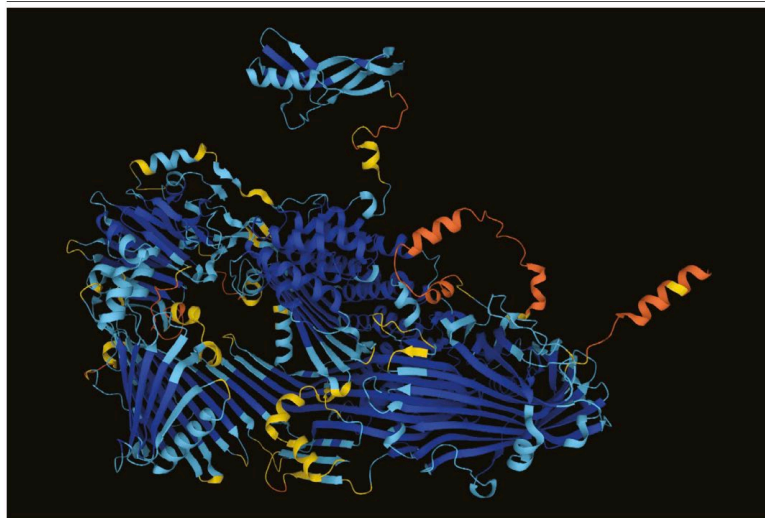
## ‘New era in digital biology’: AI reveals structures of nearly all known proteins

Advance from DeepMind’s AlphaFold software could revolutionize biology and medicine

29 JUL 2022 • 11:25 AM • BY JOHN TRAVIS

The world this week

## News in focus



The structure of the vitellogenin protein — a precursor of egg yolk — as predicted by the AlphaFold tool.

## ‘THE ENTIRE PROTEIN UNIVERSE’: AI PREDICTS SHAPE OF NEARLY EVERY KNOWN PROTEIN

DeepMind’s AlphaFold tool has determined around 200 million protein structures, which are now available to scientists in a database.

By Ewen Callaway

available for free in a database set up by DeepMind—the London-based AI company, owned

The 3D shape, or structure, of a protein is what determines its function in cells. Most



# NESG NMR, X-ray Pairs

NMR

AF

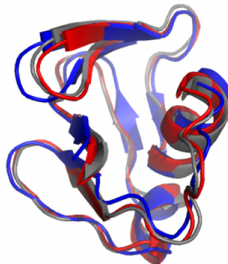
X-ray

CtR107



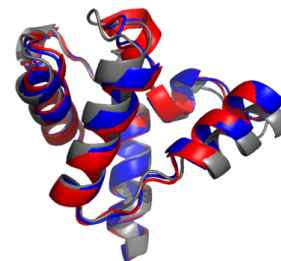
CtR107	2KCU	AF	X-ray
NMR 2KCU	2.11 Å	64.7	67.0
AF	3.89 Å	0.25 Å	97.3
X-ray 3EOH	3.62 Å	1.09 Å	----

GmR137



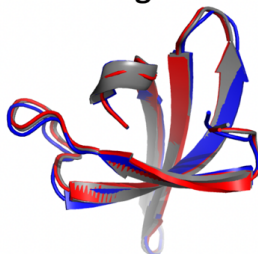
GmR137	2K5P	AF	X-ray
NMR 2K5P	0.78 Å	82.5	81.1
AF	1.52 Å	0.21 Å	98.8
X-ray 3CWI	1.59 Å	0.64 Å	----

RpR324



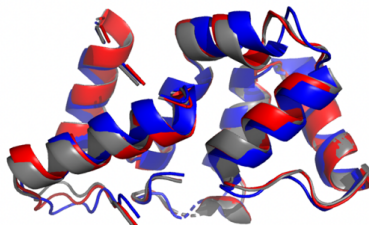
RpR324	7TZD	2LPK*	AF	X-ray
NMR 7TZD	0.54 Å	92.6	92.9	90.3
NMR 2LPK*	1.10 Å	0.45 Å	99.4	86.4
AF	1.13 Å	0.59 Å	0.27 Å	86.6
X-ray 3LMO	1.22 Å	1.68 Å	1.74 Å	----

SgR42



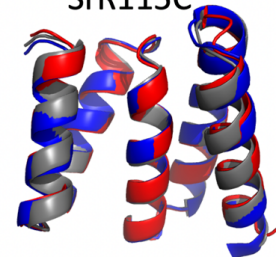
SgR42	2JZ2	AF	X-ray
NMR 2JZ2	0.66 Å	94.2	93.3
AF	0.86 Å	0.09 Å	99.6
X-ray 3C4S	0.97 Å	0.48 Å	----

SgR209C



SgR209C	2L06	7TZ8*	AF	X-ray
NMR 2L06	0.84 Å	82.7	82.7	82.2
NMR 7TZ8*	1.42 Å	0.94 Å	91.3	88.4
AF	1.51 Å	1.16 Å	0.11 Å	99.2
X-ray 3OSJ	1.53 Å	1.32 Å	0.52 Å	----

SrR115C



SrR115C	2KCL	2KCV*	AF	X-ray
NMR 2KCL	0.47 Å	92.9	93.4	87.4
NMR 2KCV*	1.03 Å	0.43 Å	95.6	95.8
AF	1.27 Å	1.28 Å	0.14 Å	94.7
X-ray 3MA5	1.35 Å	0.78 Å	1.13 Å	----



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Contents lists available at [ScienceDirect](https://www.sciencedirect.com)

# Journal of Magnetic Resonance

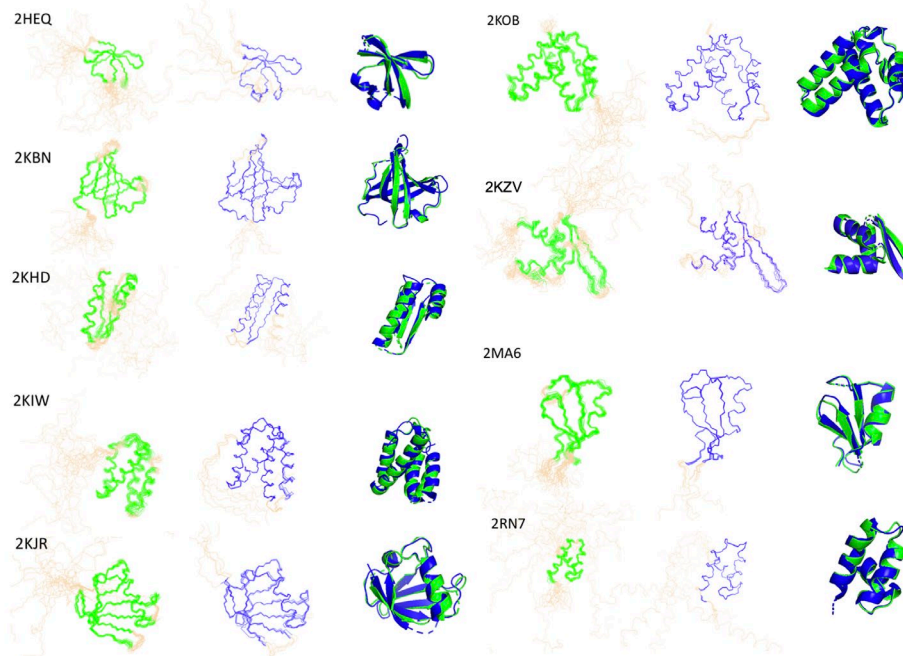
journal homepage: [www.elsevier.com/locate/jmr](http://www.elsevier.com/locate/jmr)



## Blind assessment of monomeric AlphaFold2 protein structure models with experimental NMR data



Ethan H. Li<sup>a</sup>, Laura E. Spaman<sup>a</sup>, Roberto Tejero<sup>a</sup>, Yuanpeng Janet Huang<sup>a</sup>, Theresa A. Ramelot<sup>a</sup>, Keith J. Fraga<sup>a</sup>, James H. Prestegard<sup>b</sup>, Michael A. Kennedy<sup>c</sup>, Gaetano T. Montelione<sup>a,\*</sup>





*Homo sapiens*

# Thrombomodulin

## AlphaFold structure prediction

EMBL-EBI



UniProt



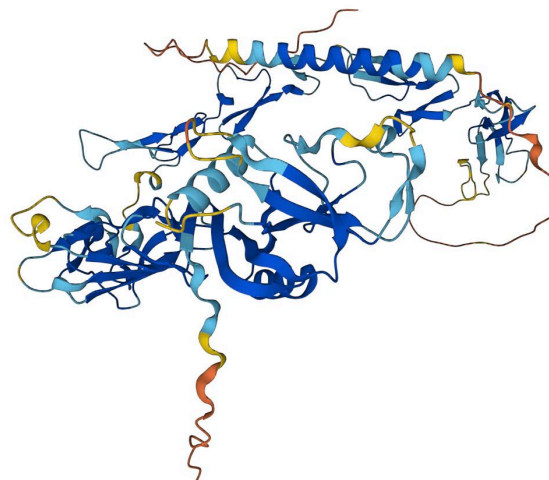
### 3D viewer

#### Model Confidence:

- Very high (pLDDT > 90)
- Confident (90 > pLDDT > 70)
- Low (70 > pLDDT > 50)
- Very low (pLDDT < 50)

AlphaFold produces a per-residue confidence score (pLDDT) between 0 and 100. Some regions below 50 pLDDT may be unstructured in isolation.

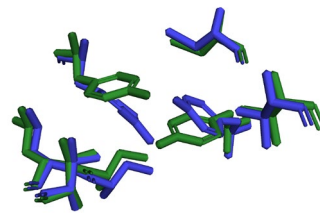
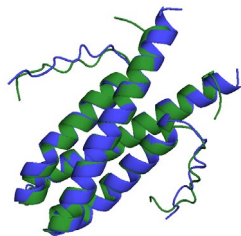
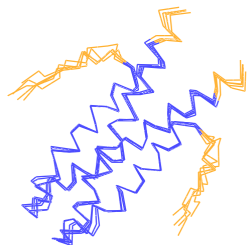
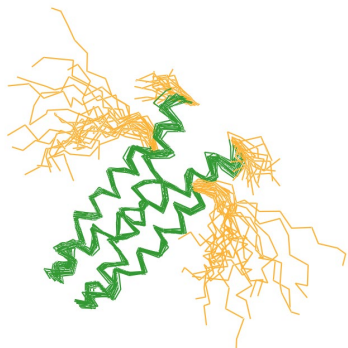
```
Sequence of AF-P07204-F1 Chain 1: Thrombomodulin A
1 11 21 31 41 51 61 71 81 91 101 111 121
MLGVLVLGALALAGLGFPAPEPQGGGQVVEHDCFALYPGPATFLNASQICDGLRGHLMTVRSVAADVISLLNGDGGVRRRLWIGLQLPPGCGDKRRLGFLRGFQVVTGDNNTSYSRWAR
131 141 151 161 171 181 191 201 211 221 231 241
LDLNGAPLCGLCVAVSAAEATVPSEPIWEEQQCEVKADGFLCEPHFATCRPLAVEPGAAAAVSIYGTPPAARGADFQALPVGSSAAVPLGLQIMCTAPPQAVQGHWAREAPGAWDCSVE
251 261 271 281 291 301 311 321 331 341 351 361 371
NGGCEHACNAIPGAPRCQCPAGAALQADGRSCTASATQSCNDLCEHFCVFNPDQPGSYSCMCTGYRLAADQHRCEVDVDDCILEPSPQRCVNTQGGFECHCYPNYDLVDGECVEPVDPCFRA
```



Jumper, J et al. Highly accurate protein structure prediction with AlphaFold. Nature (2021).

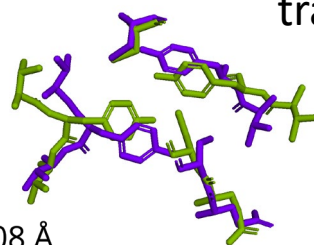
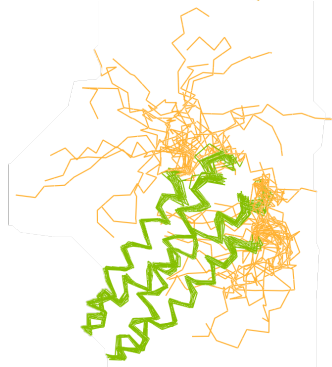
Varadi, M et al. AlphaFold Protein Structure Database: massively expanding the structural coverage of protein-sequence space with high-accuracy models. Nucleic Acids Research (2021).

# DOC1 and DOC1R – Deleted in Oral Cancer. CDK2 modulating proteins



RMSD = 0.65 Å  
GDT\_TS = 0.99

Not used  
in AF  
training



RMSD = 1.08 Å  
GDT\_TS = 0.94

L Spaman

Protein	Method	<DP> <sup>a</sup>	DP <sub>avg</sub> <sup>a</sup>	R <sub>avg</sub> <sup>a</sup>	P <sub>avg</sub> <sup>a</sup>	F <sub>avg</sub> <sup>a</sup>	ProCheck - bb <sup>b</sup>	ProCheck - all <sup>b</sup>	Mol Probity <sup>b</sup>	Rama-chandran Statistics <sup>c</sup>
CDK2AP1	NMR	0.77	0.71	0.97	0.90	0.93	+2.44	+2.48	-0.84	99.3/0.6
	AF	0.69	0.68	0.95	0.86	0.90	+1.85	+1.89	+1.53	91.5/8.5
CDK2AP2	NMR	0.80	0.72	0.96	0.92	0.94	+2.36	+1.83	+0.08	98.9/1.1
	AF	0.77	0.73	0.96	0.90	0.93	+1.89	+1.42	+0.41	89.4/10.6

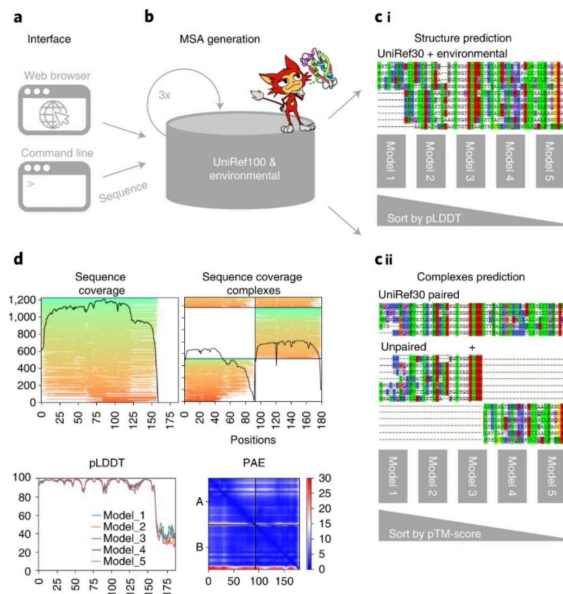
## ColabFold: making protein folding accessible to all

[Milot Mirdita](#) , [Konstantin Schütze](#), [Yoshitaka Moriwaiki](#), [Lim Heo](#), [Sergey Ovchinnikov](#)  & [Martin Steinegger](#) 

*Nature Methods* **19**, 679–682 (2022) | [Cite this article](#)

Web based  
AF2 modeling

**Fig. 1: Schematic diagram of ColabFold.**



**a, b**, ColabFold has a web and a command line interface (**a**) that send FASTA input sequence(s) to an MMSeqs2 server (**b**) searching two databases, UniRef100 and a database of environmental sequences,

Democratization  
of  
Structural  
Biology

# Evolutionary-scale prediction of atomic-level protein structure with a language model

ZEMING LIN , HALIL AKIN , ROSHAN RAO , BRIAN HIE , ZHONGKAI ZHU, WENTING LU, NIKITA SMETANIN, ROBERT VERKUIJ , ORI KABELI , [..], AND

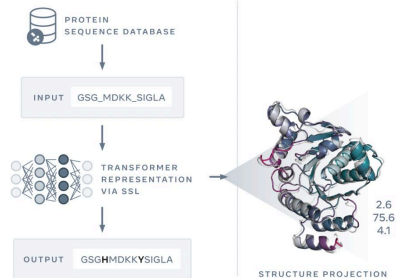
ALEXANDER RIVES 

+5 authors

[Authors Info & Affiliations](#)

Protein language modeling

Pretraining self-supervision on sequences only. → Structure emerges in the internal representations of the network from the self-supervision.



The ESM-2 language model is trained to predict amino acids that have been masked out of sequences across evolution. We discovered that, as a result of this training, information about the protein's structure emerges in the internal states of the model. This is surprising because the model has been trained only on sequences.

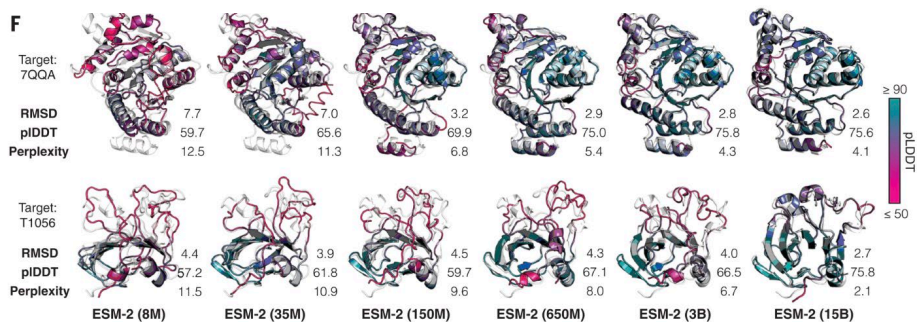


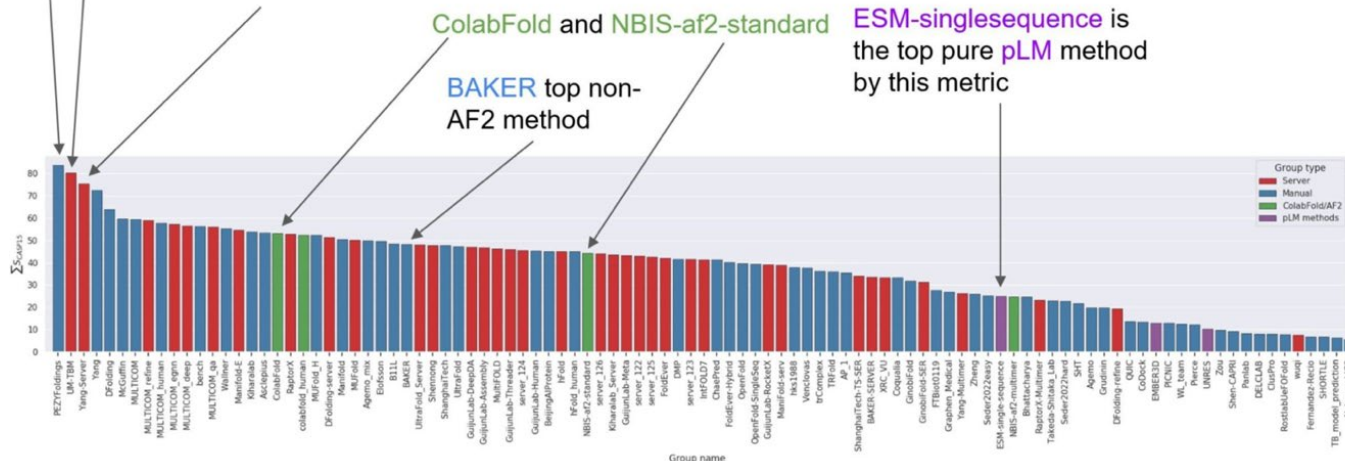
Fig. 1. Emergence of structure when scaling language models to 15 billion parameters.

#1 PEZYFoldings AF2-based. Diverse MSAs.  
Custom, fine-tuned AF2 refinement

#2 UM-TBM Diverse MSAs. Threading then AF2  
predictions guide I-TASSER REMC

#3 Yang-Server Diverse MSAs. AF2  
predictions fed to trRosettaX2

# The CASP15 rankings





# OpenFold

Democratizing AI for Biology

## Why OpenFold?



### Open Source

An open-source project that can be used and improved by academics and companies alike.



### Weights Available

Utilize the existing pre-trained weights to get quickly get started fine-tuning your model.



### Permissive License

A permissively licensed model that allows commercial and non-commercial use.



### Training Pipeline

Provides the tools used to train the model under the same license.



### Optimized for Performance

Optimized performance for use on state-of-the-art and widely available GPUs.



### PyTorch-Based

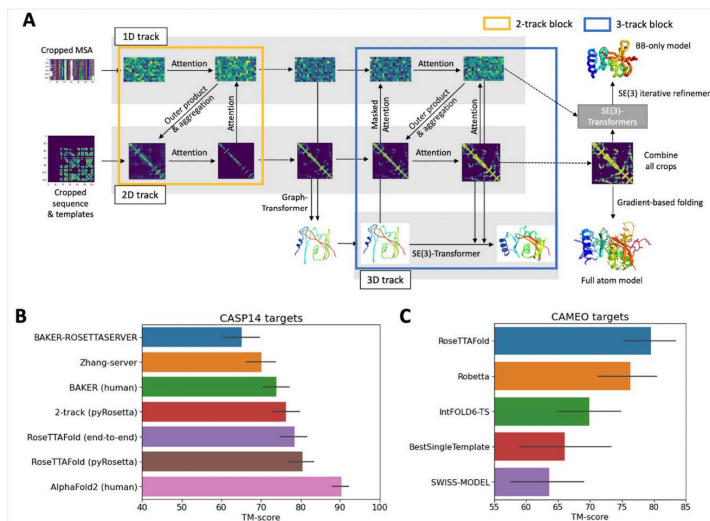
A supercomputer scale, distributed training, PyTorch-based training framework



Cite as: M. Baek *et al.*, *Science*  
10.1126/science.abj8754 (2021).

# Accurate prediction of protein structures and interactions using a three-track neural network

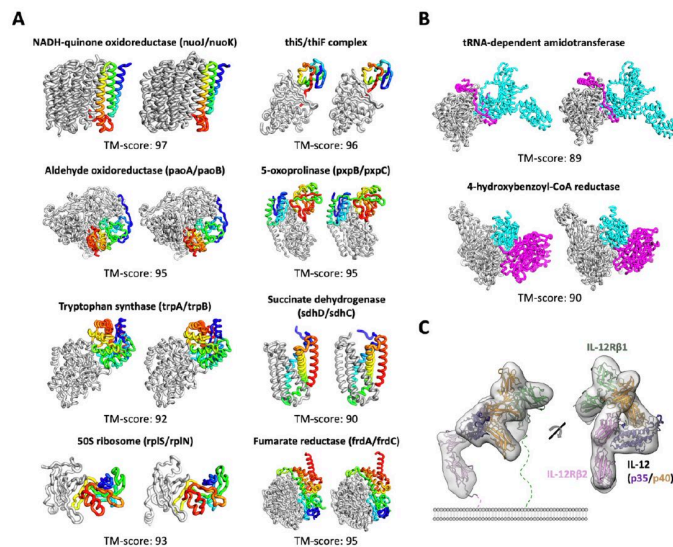
Minkyung Baek<sup>1,2</sup>, Frank DiMaio<sup>1,2</sup>, Ivan Anishchenko<sup>1,2</sup>, Justas Dauparas<sup>1,2</sup>, Sergey Ovchinnikov<sup>3,4</sup>, Gyu Rie Lee<sup>1,2</sup>, Jue Wang<sup>1,2</sup>, Qian Cong<sup>5,6</sup>, Lisa N. Kinch<sup>7</sup>, R. Dustin Schaeffer<sup>6</sup>, Claudia Millán<sup>8</sup>, Hahnbeom Park<sup>1,2</sup>, Carson Adams<sup>1,2</sup>, Caleb R. Glassman<sup>9,10</sup>, Andy DeGiovanni<sup>12</sup>, Jose H. Pereira<sup>12</sup>, Andria V. Rodrigues<sup>12</sup>, Alberdina A. van Dijk<sup>13</sup>, Ana C. Ebrecht<sup>13</sup>, Diederik J. Opperman<sup>14</sup>, Theo Sagmeister<sup>15</sup>, Christoph Buhlheller<sup>15,16</sup>, Tea Pavkov-Keller<sup>15,17</sup>, Manoj K. Rathinaswamy<sup>18</sup>, Udit Dalwadi<sup>19</sup>, Calvin K. Yip<sup>19</sup>, John E. Burke<sup>18</sup>, K. Christopher Garcia<sup>9,10,11,20</sup>, Nick V. Grishin<sup>6,21,7</sup>, Paul D. Adams<sup>12,22</sup>, Randy J. Read<sup>8</sup>, David Baker<sup>1,2,23\*</sup>



Cite as: M. Baek *et al.*, *Science*  
10.1126/science.abj8754 (2021).

# Accurate prediction of protein structures and interactions using a three-track neural network

## Protein-Protein Complexes



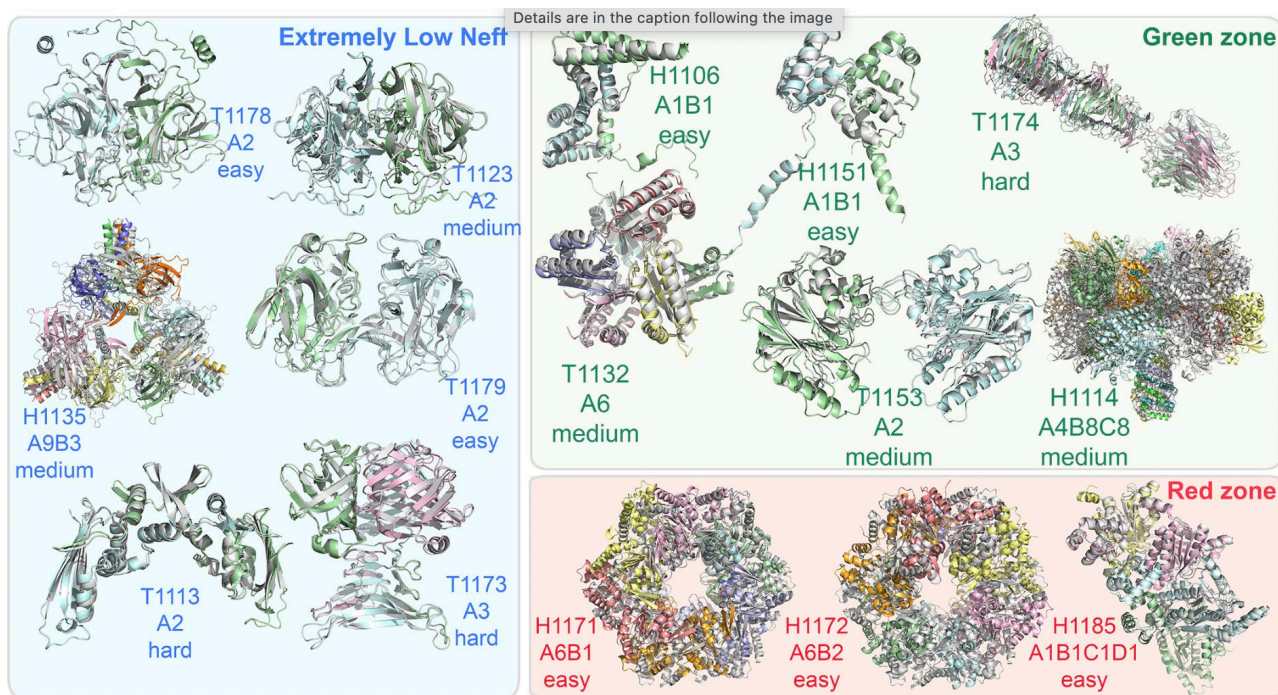
**Fig. 4. Complex structure prediction using RoseTTAFold.** (A and B) Prediction of structures of *E. coli* protein complexes from sequence information. Experimentally determined structures are on the left, RoseTTAFold models, on the right; the TM-scores below indicate the extent of structural similarity. (A) Two chain complexes. The first subunit is colored in gray, and the second subunit is colored in a rainbow from blue (N-terminal) to red (C-terminal). (B) Three chain complexes. Subunits are colored in gray, cyan, and magenta. (C) IL-12R/IL-12 complex structure generated by RoseTTAFold fits the previously published cryo-EM density (EMD-21645).



## The impact of AI-based modeling on the accuracy of protein assembly prediction: Insights from CASP15

Burcu Ozden, Andriy Kryshchak, Ezgi Karaca

First published: 20 October 2023 | <https://doi.org/10.1002/prot.26598> | Citations: 2

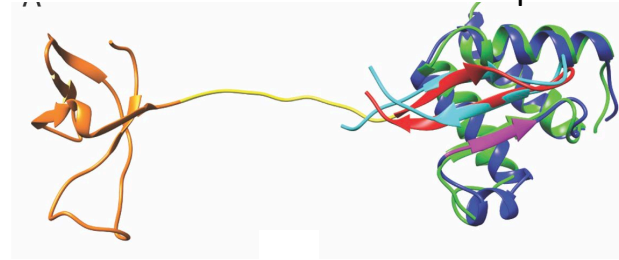




# BET Proteins: ET : Peptide Complexes

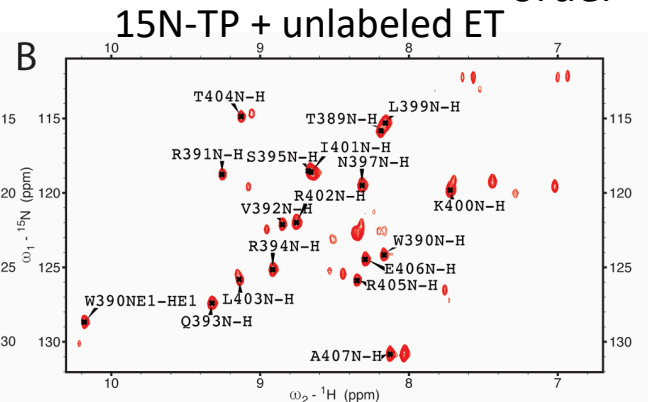
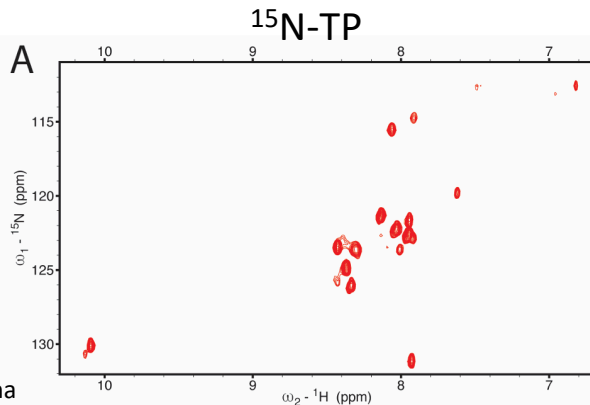
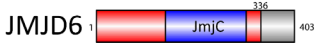
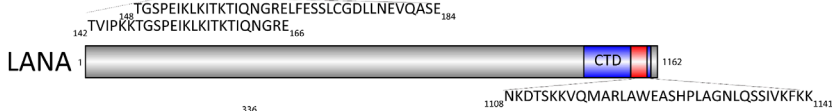
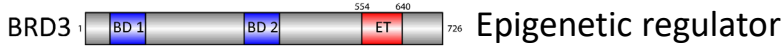
MLV IN CTD

ET – TP complex



Epigenetic regulators  
and viral proteins

Disorder ->  
order

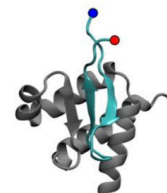


## Structure Determination of Challenging Protein–Peptide Complexes Combining NMR Chemical Shift Data and Molecular Dynamics Simulations

Arup Mondal, G.V.T. Swapna, Maria M. Lopez, Laura Klang, Jingzhou Hao, LiChung Ma,  
Monica J. Roth,\* Gaetano T. Montelione,\* and Alberto Perez\*

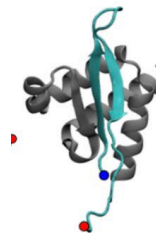
Disorder ->  
order

Experimental



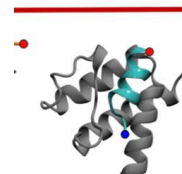
ET-TP

tight  
 $K_d$  90 nM



ET-NSD3

weak  
 $K_d \sim 250 \mu\text{M}$



ET-JMJD6

v. weak  
 $K_d \sim 950 \mu\text{M}$

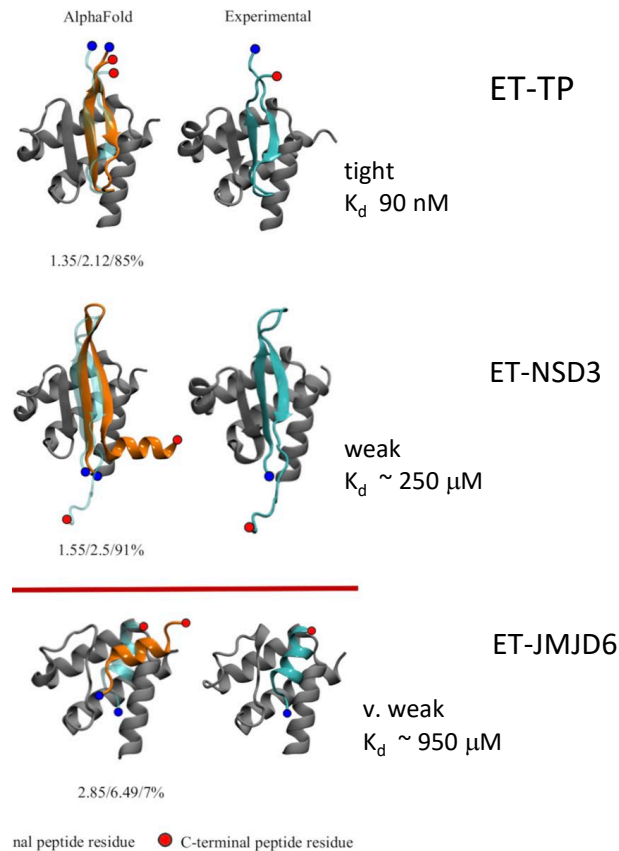
C-terminal peptide residue

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Monica J. Roth,\* Gaetano T. Montelione,\* and Alberto Perez\*

Disorder ->  
order

AF2 models of the  
peptide-receptor  
complex are an  
excellent match to  
experimental  
structures

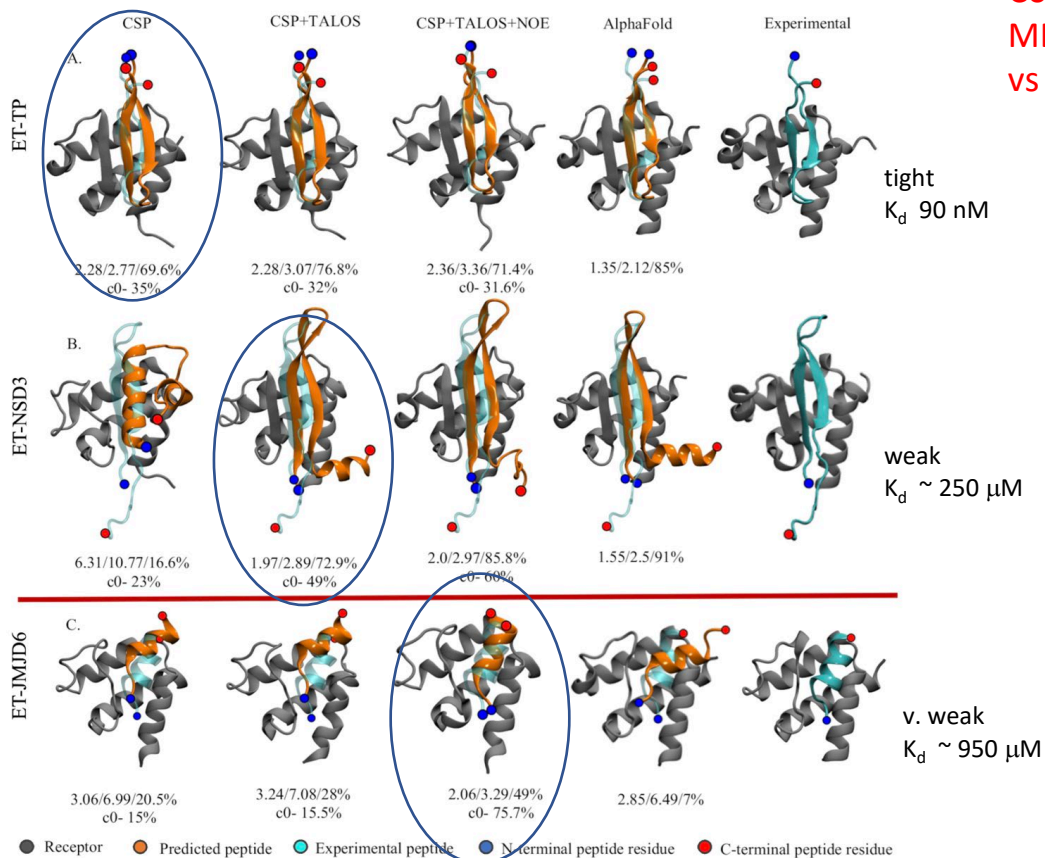


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Monica J. Roth,\* Gaetano T. Montelione,\* and Alberto Perez\*

Comparing  
MELD + NMR  
vs AlphaFold

Combining  
MD replica  
exchange  
and NMR  
chemical  
shift  
perturbation  
data through  
Bayesian  
inference



tight  
 $K_d$  90 nM

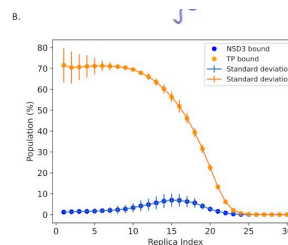
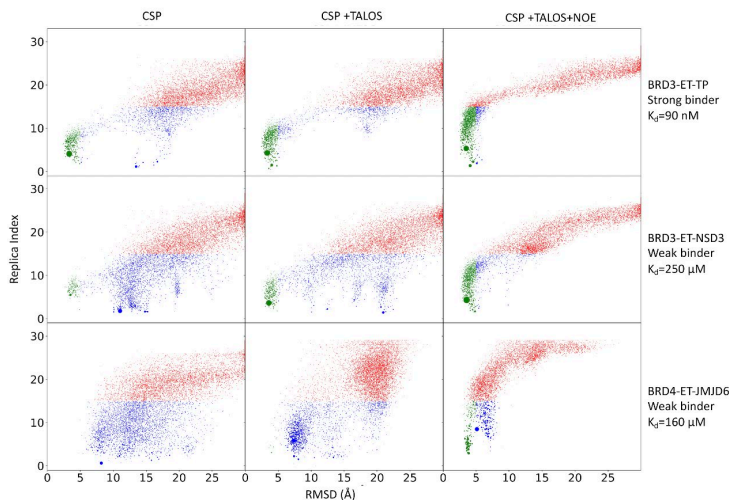
weak  
 $K_d \sim 250 \mu\text{M}$

v. weak  
 $K_d \sim 950 \mu\text{M}$

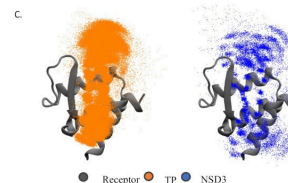


## Structure Determination of Challenging Protein–Peptide Complexes Combining NMR Chemical Shift Data and Molecular Dynamics Simulations

Arup Mondal, G.V.T. Swapna, Maria M. Lopez, Laura Klang, Jingzhou Hao, LiChung Ma, Monica J. Roth,\* Gaetano T. Montelione,\* and Alberto Perez\*



Predicting relative binding affinities



$$\Delta\Delta G_{\text{pred}} = - 2.4 \text{ kcal / mol}$$

$$\Delta\Delta G_{\text{exp}} = - 4.7 \text{ kcal / mol}$$

## AF2 for Docking -> MELD for docking free energy

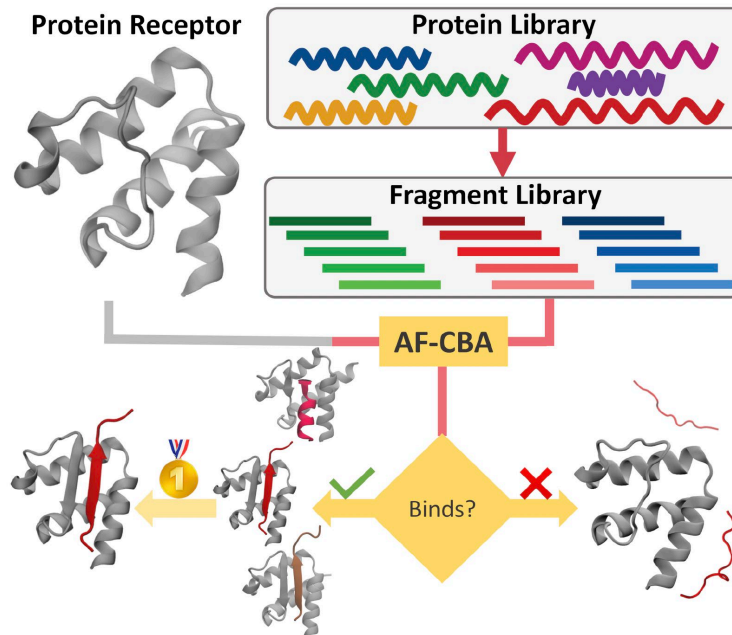
We are designing synthetic peptides that bind to ET and compete with natural ET binding proteins, by conventional de novo design and hallucination approaches.

- Antiviral or anticancer properties
- Create novel ET-protein complexes to modulate gene expression



# Sifting Through the Noise: A Computational Pipeline for Accurate Prioritization of Protein-Protein Binding Candidates in High-Throughput Protein Libraries

Arup Mondal, Bhumika Singh, Roland H. Felkner, Anna De Falco, GVT Swapna, Gaetano T. Montelione, Monica J. Roth, Alberto Perez



AF-CBA:

AlphaFold  
Competition  
Binding  
Assay

For 11 of 15 cases tested; AF-CBA identified correct region of the protein to bind to receptor, and correct experimental binding mode

A. Perez

A. Mondal

Univ of Florida

M. Roth - Rutgers

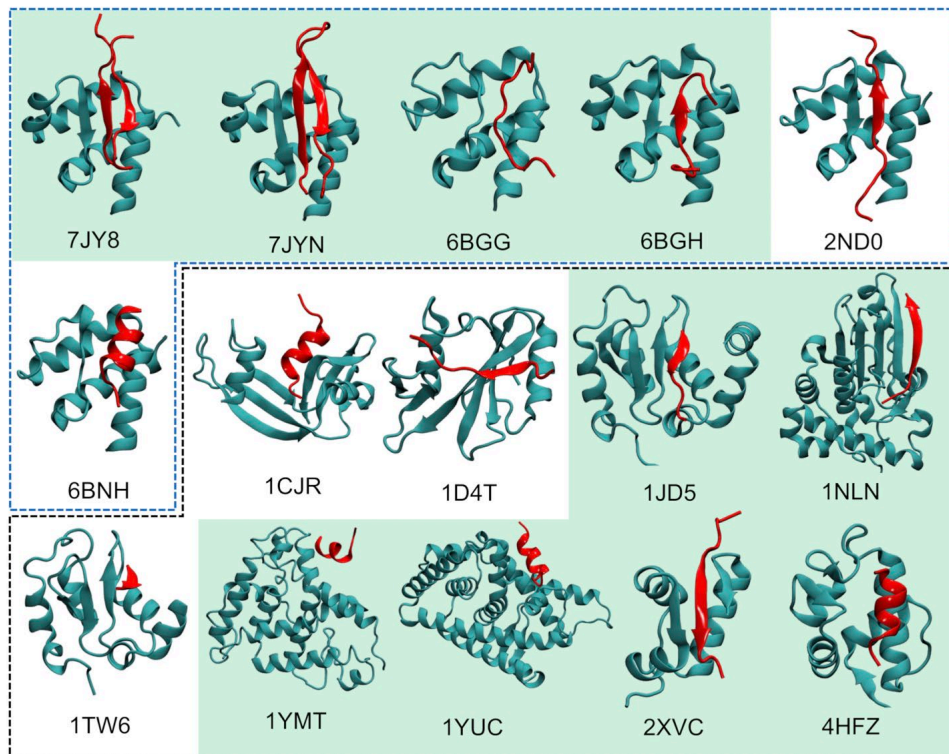
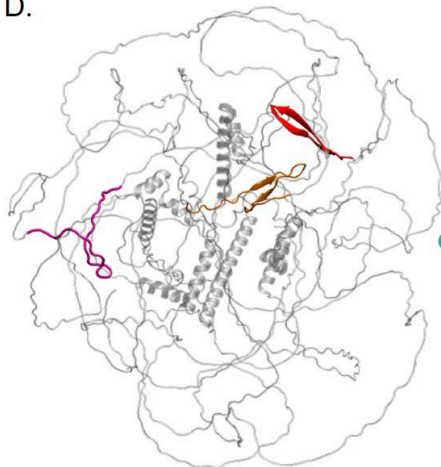
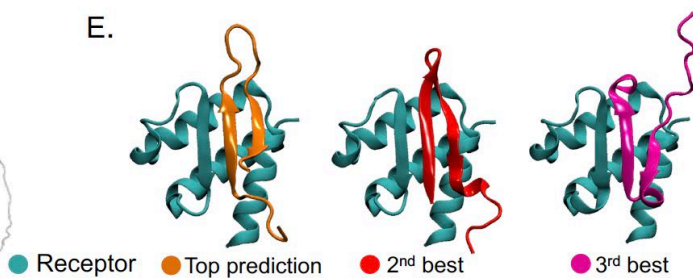


Figure 3. Test cases for AF-CBA pipeline. Experimental structures of protein complexes used in this study with their PDB code. The receptor protein is shown in cyan and peptides in red. The blue dashed line denotes systems involving the ET receptor (Proof of concept 1), whereas those inside the black dashed line show transferability to other systems (Proof of concept 2). Successful cases are highlighted with green background, new insights and limitations are discussed in the text for those with a white background. These representations are rendered with VMD<sup>[26]</sup>.

D.

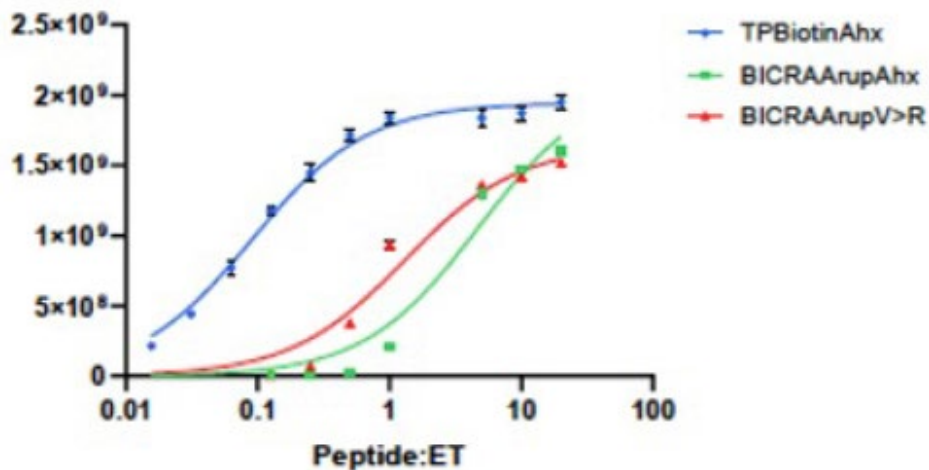


E.



Predicted  
BIRCA  
binding  
peptides with  
ET receptor

BICRA  
BET-binding  
protein  
identified by  
pull down



# NESG BIOMEDICAL THEME PROJECT

## Human Cancer Protein Interaction Network (HCPIN)

### KEGG Pathways

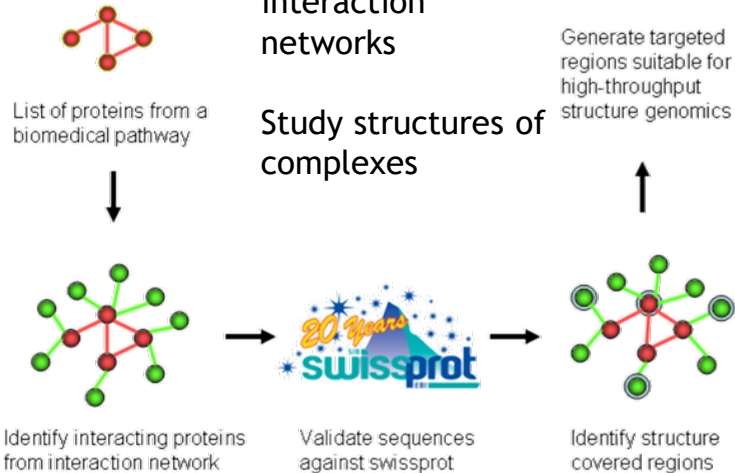
- Apoptosis
- TGF-beta
- Innate Immune Response
- EGF receptor pathway
- NF- $\kappa$ B
- JAK/Stat
- MAPK

Protein Interaction Data from HPRD: Manually curated db of protein-protein interactions based largely on literature, describing physical protein-protein interactions

Systematic structural coverage of pathways and interaction networks

Study structures of complexes

Generate targeted regions suitable for high-throughput structure genomics

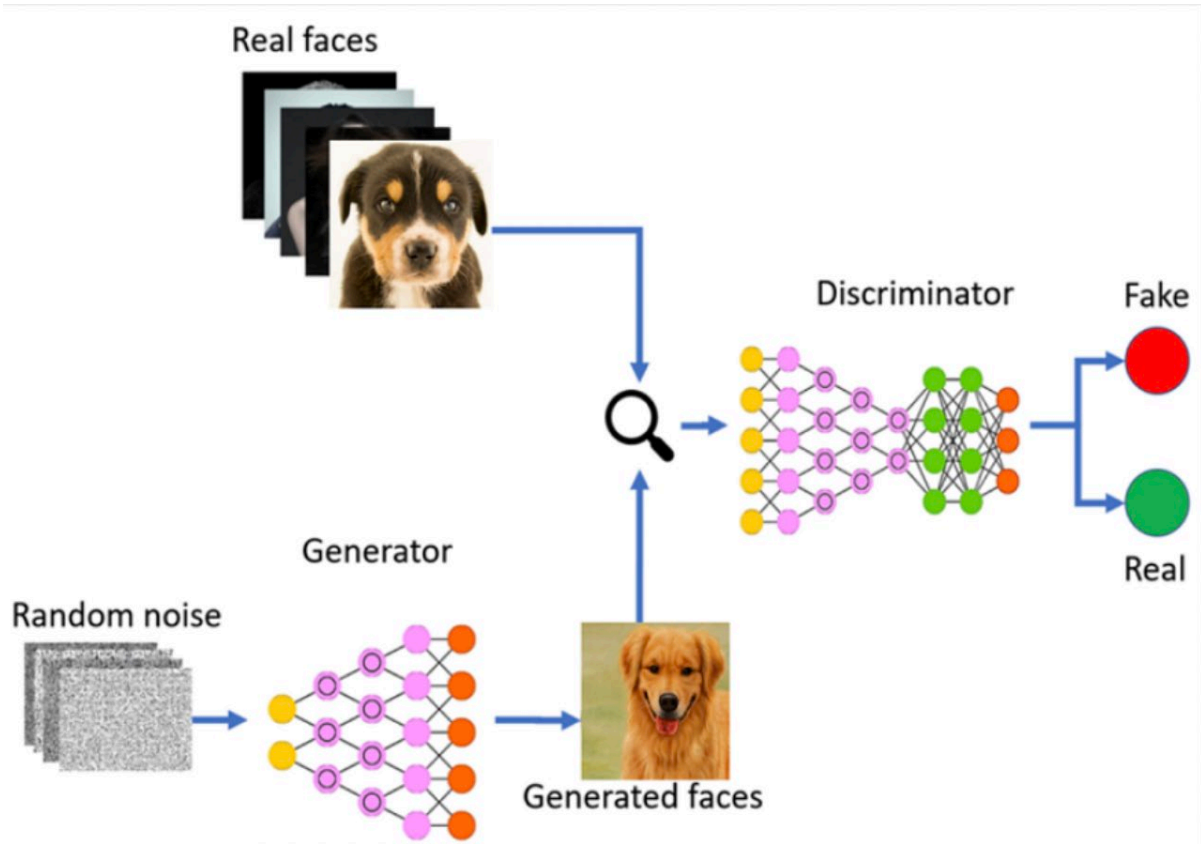


## Targeting the Human Cancer Pathway Protein Interaction Network by Structural Genomics\*

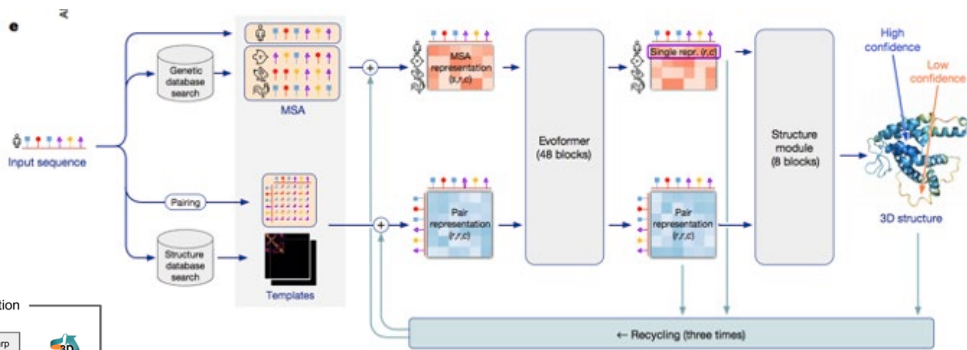
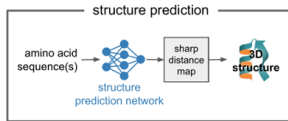
Yuanpeng Janet Huang<sup>‡</sup>, Dehua Hang<sup>‡</sup>, Long Jason Lu<sup>§||</sup>, Liang Tong<sup>||</sup>, Mark B. Gerstein<sup>§\*\*</sup>, and Gaetano T. Montelione<sup>‡ ††</sup>



# Generative Adversarial Networks (GANs) - Hallucination







Jumper et al Nature 2021

nature

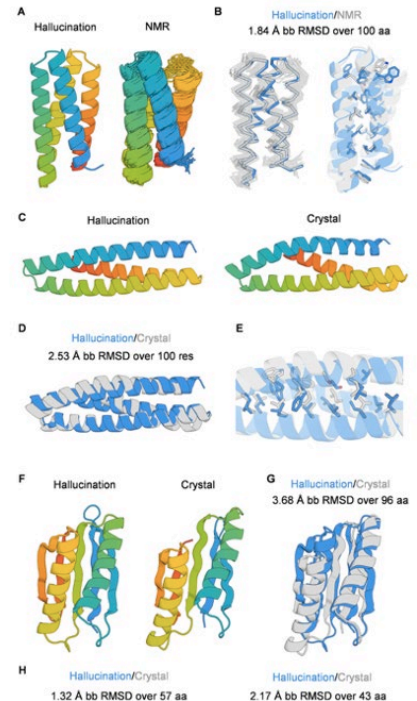
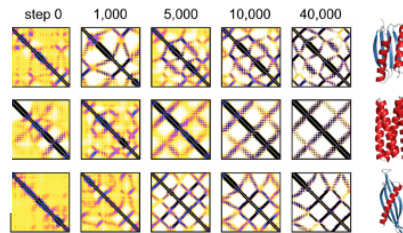
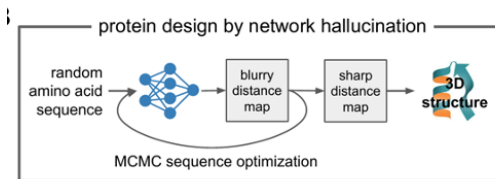
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Article | Published: 01 December 2021

## De novo protein design by deep network hallucination

Ivan Anishchenko, Samuel J. Pellock, Tamuka M. Chidyausiku, Theresa A. Ramelot, Sergey Ovchinnikov, Jingzhou Hao, Khushboo Bafna, Christoffer Norn, Alex Kang, Asim K. Bera, Frank DiMaio, Lauren Carter, Cameron M. Chow, Gaetano T. Montelione & David Baker

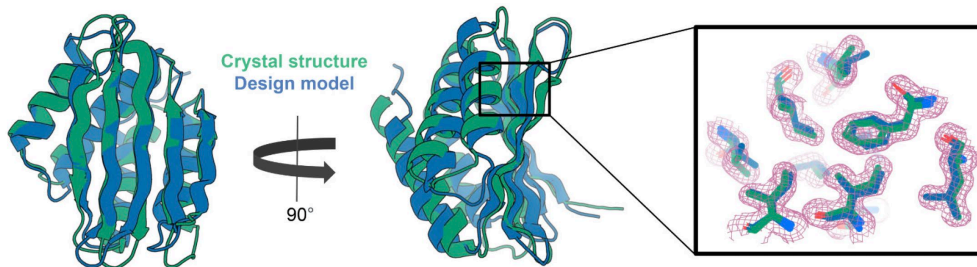
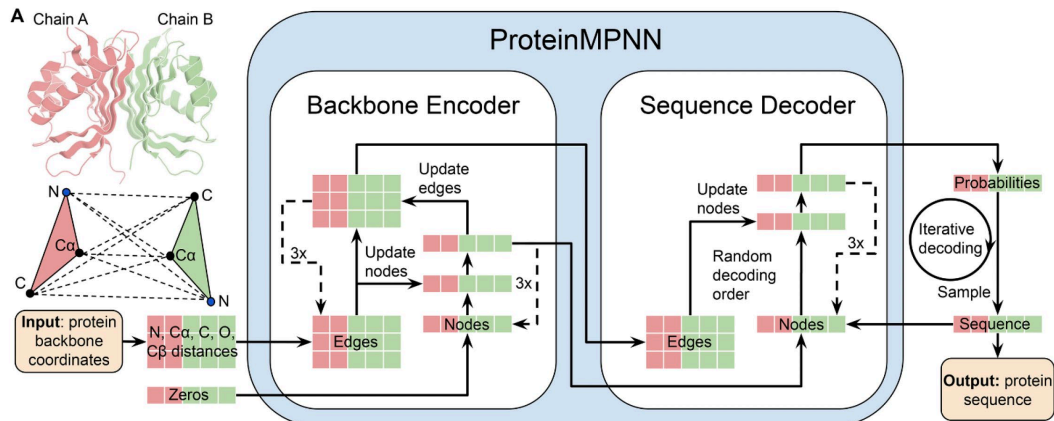


T Ramelot

Cite as: J. Dauparas *et al.*, *Science*  
10.1126/science.add2187 (2022).

# Robust deep learning–based protein sequence design using ProteinMPNN

J. Dauparas<sup>1,2</sup>, I. Anishchenko<sup>1,2</sup>, N. Bennett<sup>1,2,3</sup>, H. Bai<sup>1,2,4</sup>, R. J. Ragotte<sup>1,2</sup>, L. F. Milles<sup>1,2</sup>, B. I. M. Wicky<sup>1,2</sup>, A. Courbet<sup>1,2,4</sup>, R. J. de Haas<sup>5</sup>, N. Bethel<sup>1,2,4</sup>, P. J. Y. Leung<sup>1,2,3</sup>, T. F. Huddy<sup>1,2</sup>, S. Pellock<sup>1,2</sup>, D. Tischer<sup>1,2</sup>, F. Chan<sup>1,2</sup>, B. Koepnick<sup>1,2</sup>, H. Nguyen<sup>1,2</sup>, A. Kang<sup>1,2</sup>, B. Sankaran<sup>6</sup>, A. K. Bera<sup>1,2</sup>, N. P. King<sup>1,2</sup>, D. Baker<sup>1,2,4\*</sup>





# De novo design of protein structure and function with RFdiffusion

<https://doi.org/10.1038/s41586-023-06415-8>

Received: 14 December 2022

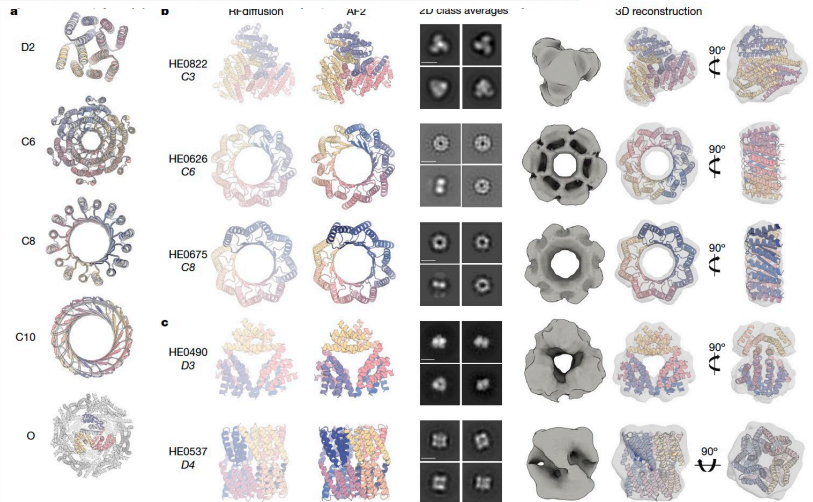
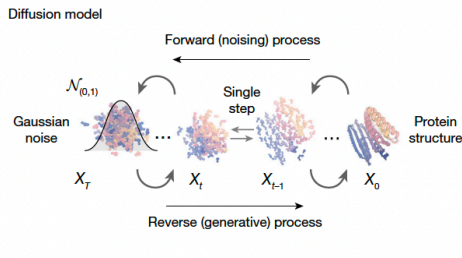
Accepted: 7 July 2023

Published online: 11 July 2023

Open access



Joseph L. Watson<sup>1,2,15</sup>, David Juergens<sup>1,2,3,15</sup>, Nathaniel R. Bennett<sup>1,2,3,15</sup>, Brian L. Trippe<sup>2,4,5,15</sup>, Jason Yim<sup>2,6,15</sup>, Helen E. Eisenach<sup>1,2,15</sup>, Woody Ahern<sup>1,2,7,15</sup>, Andrew J. Borst<sup>1,2</sup>, Robert J. Ragotte<sup>1,2</sup>, Lukas F. Milles<sup>1,2</sup>, Basile I. M. Wicky<sup>1,2</sup>, Nikita Hanikel<sup>1,2</sup>, Samuel J. Pellock<sup>1,2</sup>, Alexis Courbet<sup>1,2,8</sup>, William Sheffler<sup>1,2</sup>, Jue Wang<sup>1,2</sup>, Preetham Venkatesh<sup>1,2,9</sup>, Isaac Sappington<sup>1,2,9</sup>, Susana Vázquez Torres<sup>1,2,9</sup>, Anna Lauko<sup>1,2,9</sup>, Valentin De Bortoli<sup>8</sup>, Emile Mathieu<sup>10</sup>, Sergey Ovchinnikov<sup>11,12</sup>, Regina Barzilay<sup>6</sup>, Tommi S. Jaakkola<sup>6</sup>, Frank DiMaio<sup>1,2</sup>, Minkyung Baek<sup>13</sup> & David Baker<sup>1,2,14</sup>✉



# De novo design of protein structure and function with RFdiffusion

<https://doi.org/10.1038/s41586-023-06415-8>

Received: 14 December 2022

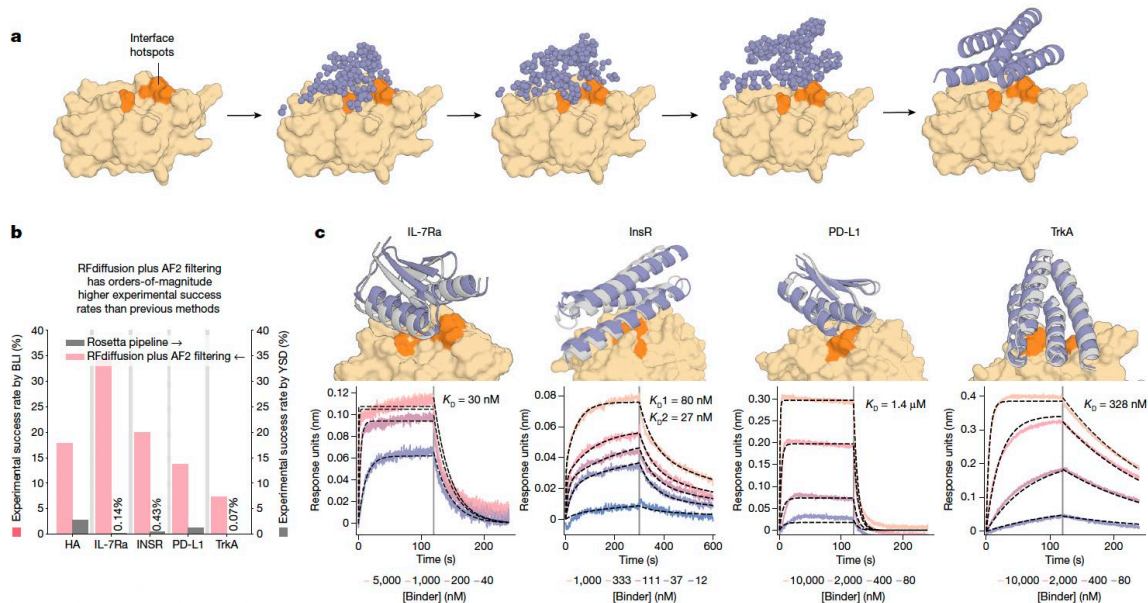
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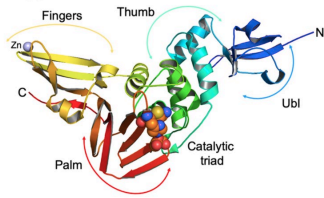
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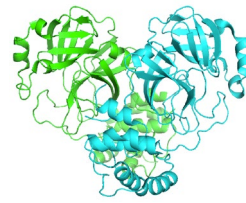
 Check for updates

Joseph L. Watson<sup>1,2,15</sup>, David Juergens<sup>1,2,3,15</sup>, Nathaniel R. Bennett<sup>1,2,3,15</sup>, Brian L. Trippe<sup>2,4,5,15</sup>, Jason Yim<sup>2,6,15</sup>, Helen E. Eisenach<sup>1,2,15</sup>, Woody Ahern<sup>1,2,7,15</sup>, Andrew J. Borst<sup>1,2</sup>, Robert J. Ragotte<sup>1,2</sup>, Lukas F. Milles<sup>1,2</sup>, Basile I. M. Wicky<sup>1,2</sup>, Nikita Hanikel<sup>1,2</sup>, Samuel J. Pellock<sup>1,2</sup>, Alexis Courbet<sup>1,2,8</sup>, William Sheffler<sup>1,2</sup>, Jue Wang<sup>1,2</sup>, Preetham Venkatesh<sup>1,2,9</sup>, Isaac Sappington<sup>1,2,9</sup>, Susana Vázquez Torres<sup>1,2,9</sup>, Anna Lauko<sup>1,2,9</sup>, Valentin De Bortoli<sup>8</sup>, Emile Mathieu<sup>10</sup>, Sergey Ovchinnikov<sup>11,12</sup>, Regina Barzilay<sup>6</sup>, Tommi S. Jaakkola<sup>6</sup>, Frank DiMaio<sup>12</sup>, Minkyung Baek<sup>13</sup> & David Baker<sup>1,2,14</sup>✉





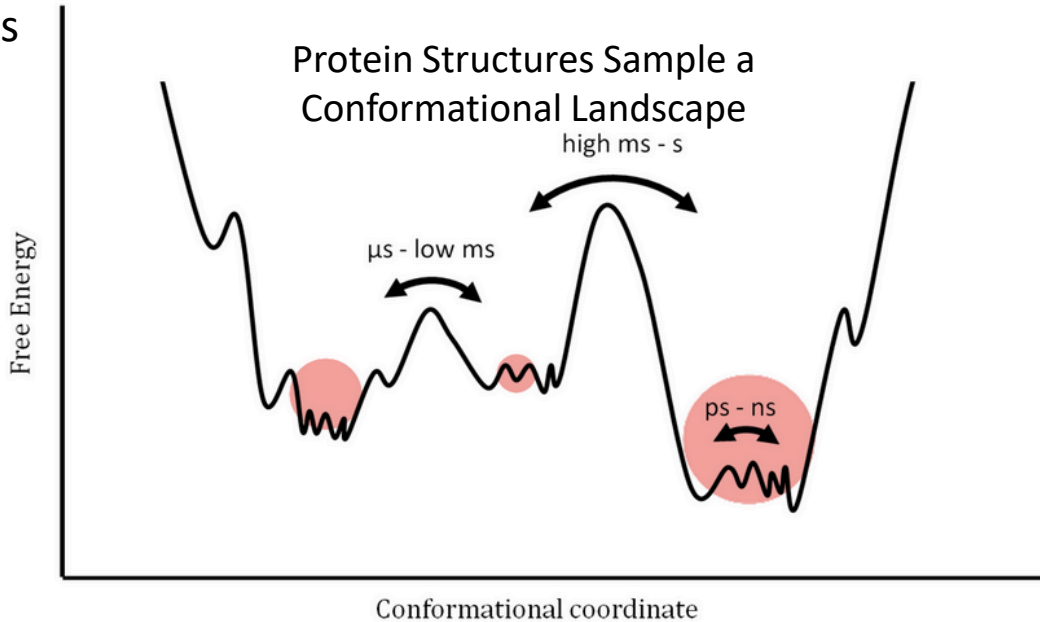
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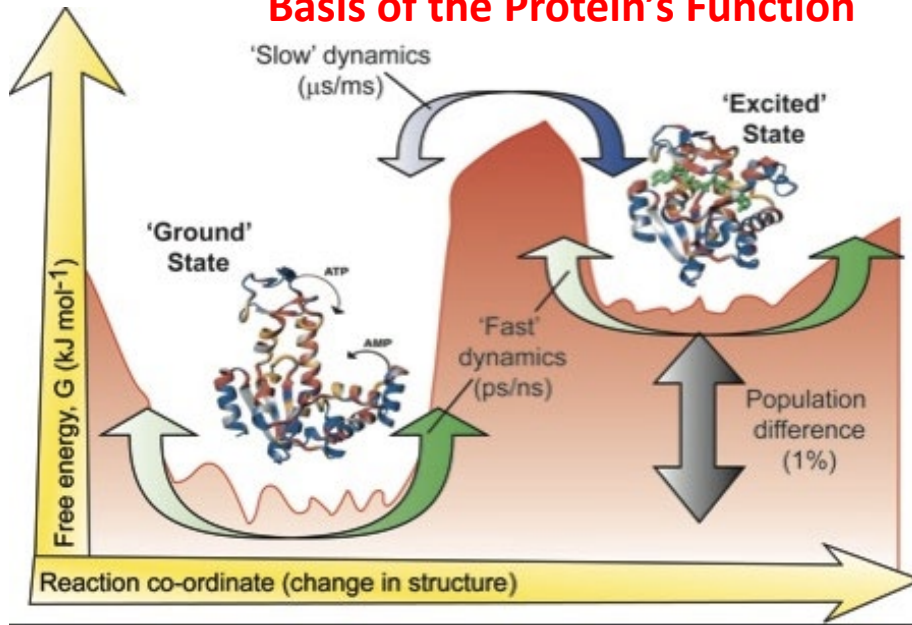
SARS CoV2 M<sup>pro</sup>

4D  
Protein  
Structures

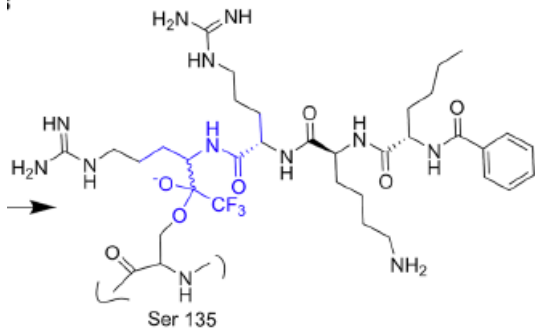
**Protein Dynamics**



## The Conformational Landscape Provides the Basis of the Protein's Function



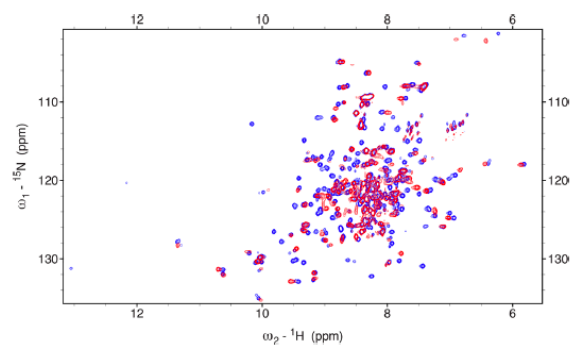
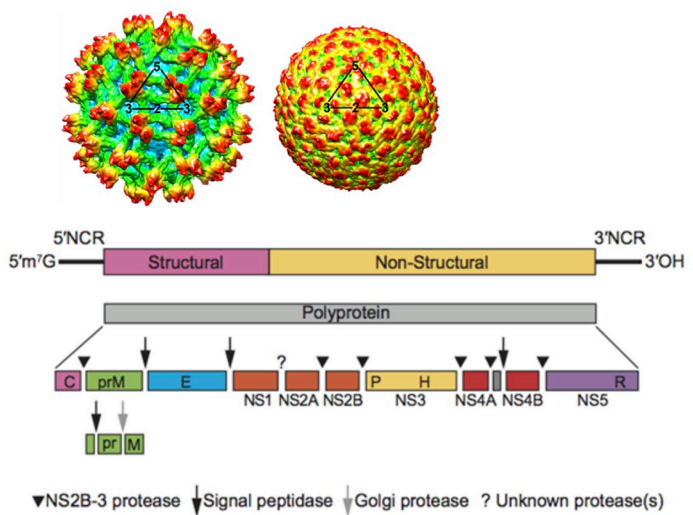
Andrew Baldwin  
research.chem.ox.ac.u  
k



Al Gibbs, Ruth Steele  
Janssen Pharma

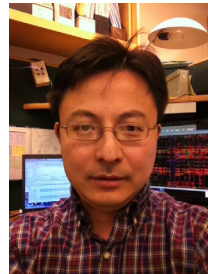
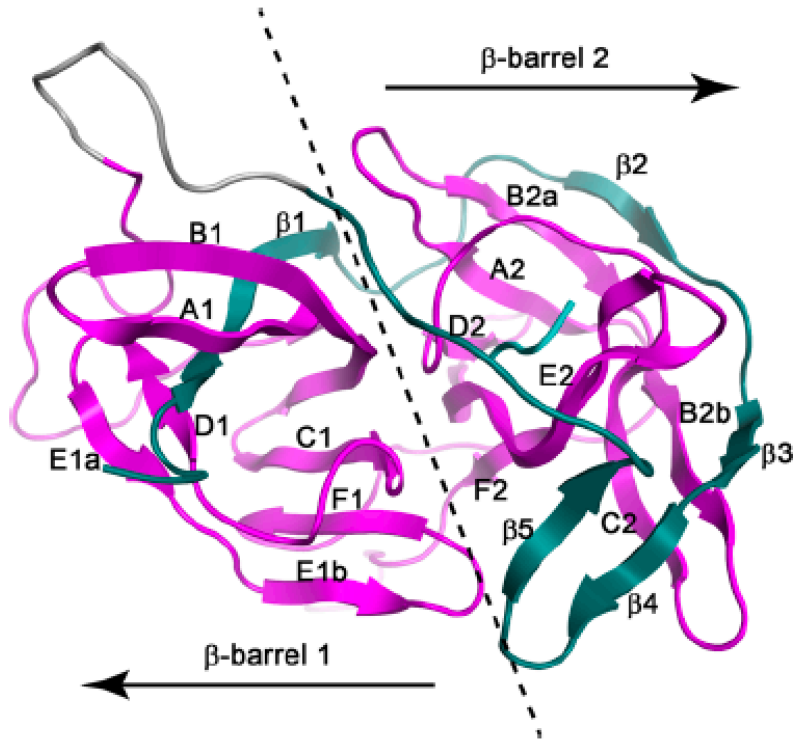
Gaohua Liu  
Nexomics Biosciences

**Figure 1** Three-dimensional cryo-electron microscopic reconstructions of immature (left) and mature (right) particles of an isolate of dengue virus (courtesy of M. Rossmann). Shown is a surface rendering of immature dengue virus at 12.5Å resolution (left) and mature DENV at 10Å resolution (right). The viruses are depicted to scale, but not colored to scale. Triangles outline one icosahedral unit.



**Figure 4.** Comparison of [ $^{15}\text{N}$ - $^1\text{H}$ ]-TROSY-HSQC spectra of apo (red) and inhibitor-bound (blue) NS2B-SN3pro. Extensive exchange broadening limits the quality of the spectrum of apo NS2B-SN3pro, while significantly more well-resolved resonances are observed in the inhibitor-bound spectra.

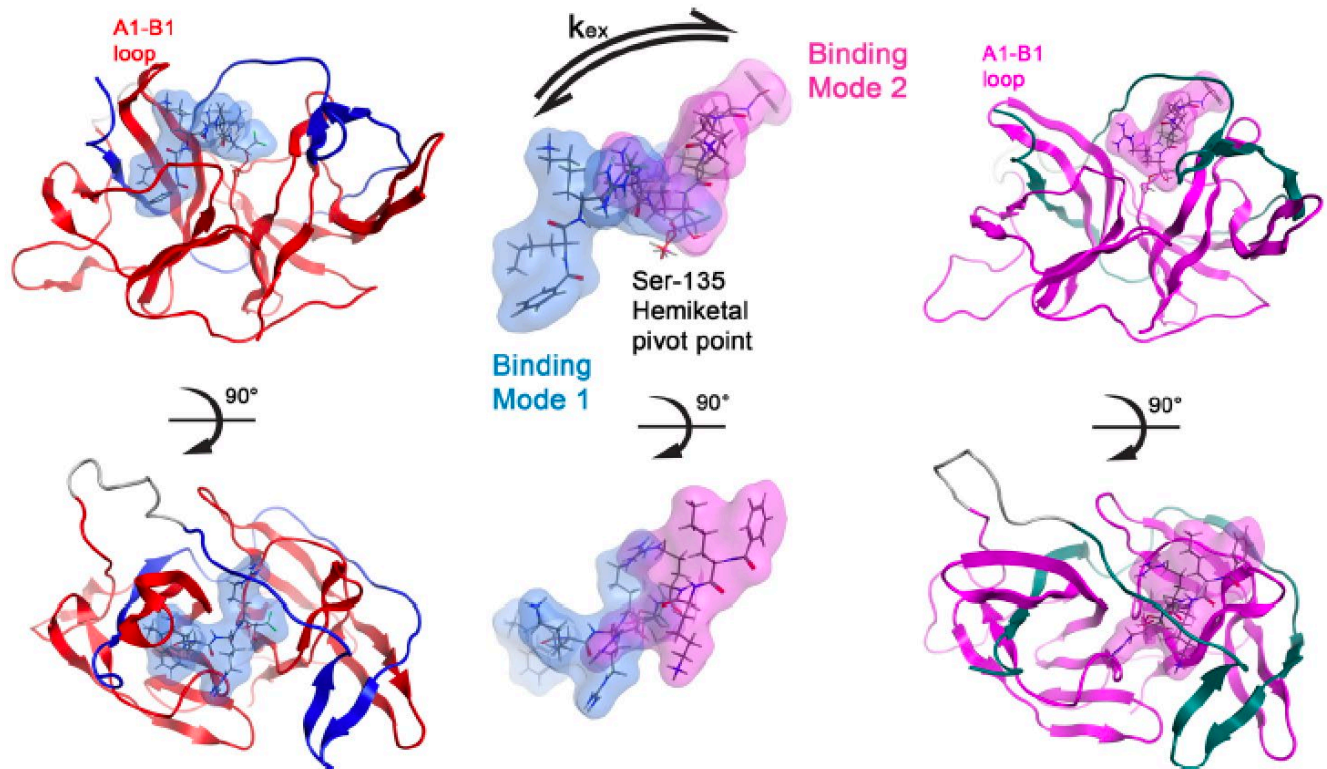
# Solution NMR Structure of DENV2-NS2B- NS3pro Protease Complex $^2\text{H}$ , $^{15}\text{N}$ , $^{13}\text{C}$ , ILVA Me Labeled



G. Liu



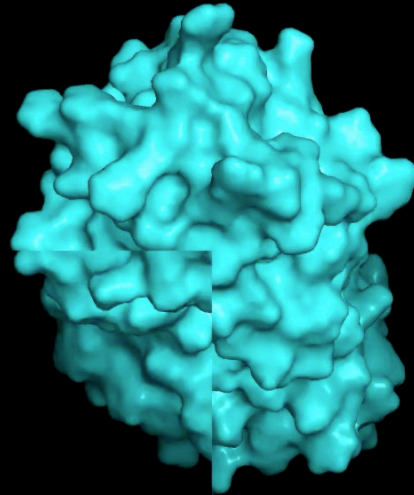
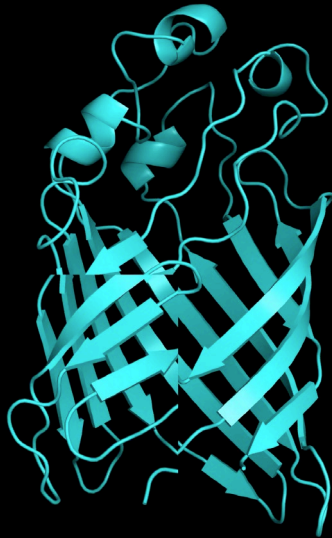
# NMR Reveals Two Non-Overlapping Inhibitor Binding Sites in DENV2-NS2B-NS3pro Protease Complex







# *Klebsiella pneumoniae* MipA in detergent micelles



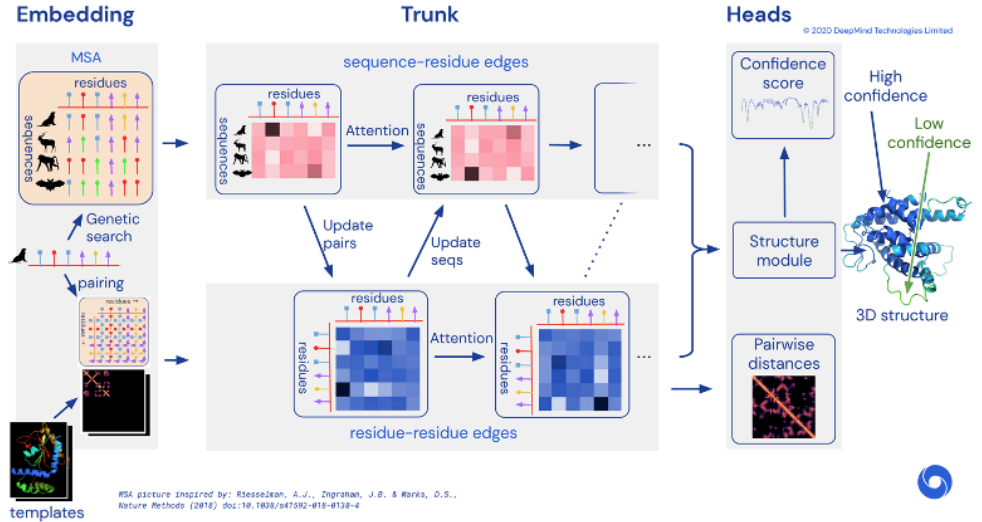
YJ Huang  
G Liu  
Y Ishida  
GVT Swapna  
S. McCallum



# Can we use AlphaFold to Predict the Multiple Conformational States of MipA?

attention-based machine learning

Use shallow MSAs to provide subsets of ECs



Jumper et al 2021

John Jumper  
CASP14 Talk Dec 2020



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Article | [Open access](#) | Published: 13 November 2023

## Predicting multiple conformations via sequence clustering and AlphaFold2

[Hannah K. Wayment-Steele](#), [Adedolapo Ojoawo](#), [Renee Otten](#), [Julia M. Apitz](#), [Warintra Pitsawong](#), [Marc Hämberger](#), [Sergey Ovchinnikov](#), [Lucy Colwell](#) & [Dorothee Kern](#)

*Nature* 625, 832–839 (2024) | [Cite this article](#)

## Bioinformatics

AFsample: improving multimer prediction with AlphaFold using massive sampling

[Björn Wallner](#)

*Bioinformatics*, Volume 39, Issue 9, September 2023, btad573,

Short Report  
Structural Biology and Molecular Biophysics

## Sampling alternative conformational states of transporters and receptors with AlphaFold2

Diego del Alamo, Davide Sala, Hassane S Mchaourab , Jens Meiler

AF-Alt

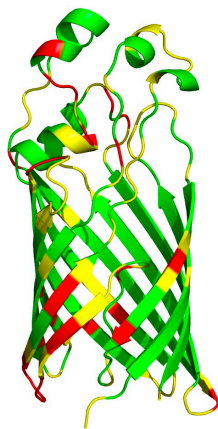
AF-cluster

AF-sample

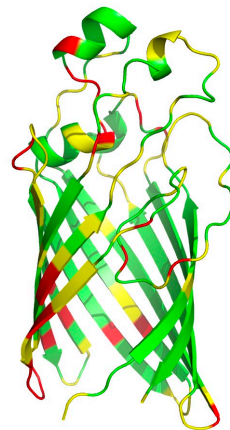
# AF\_ALT generates three clusters of models structural variation in the strand $\beta_2$ , $\beta_3$ , $\beta_4$ , $\beta_5$



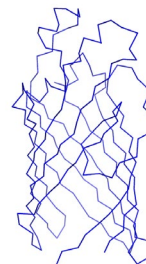
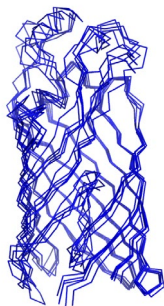
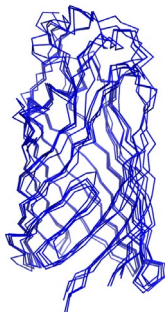
AF\_ALT\_1  
cluster 1



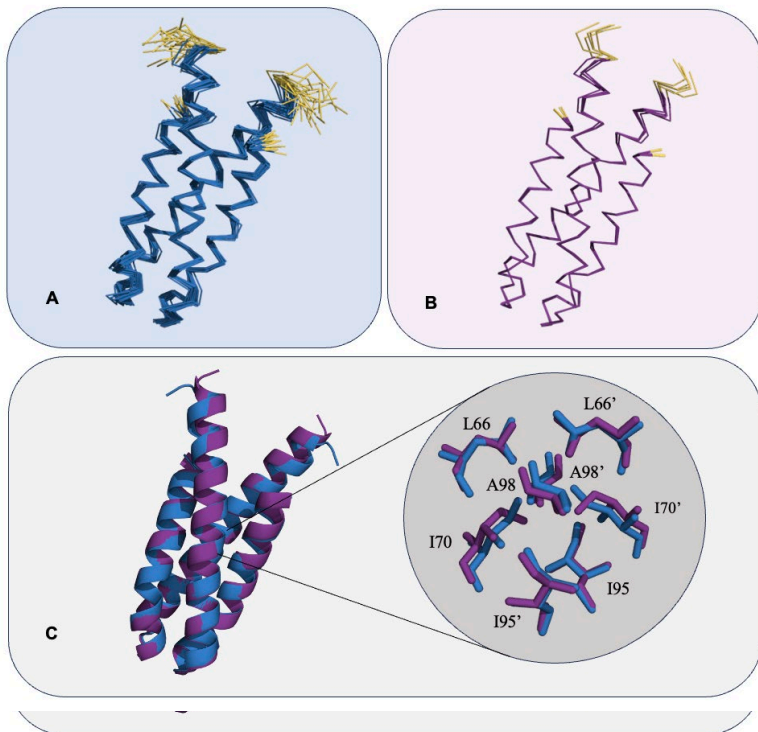
AF-ALT\_2  
cluster 2



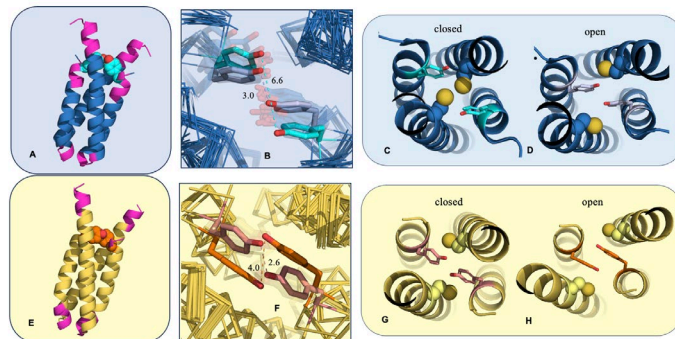
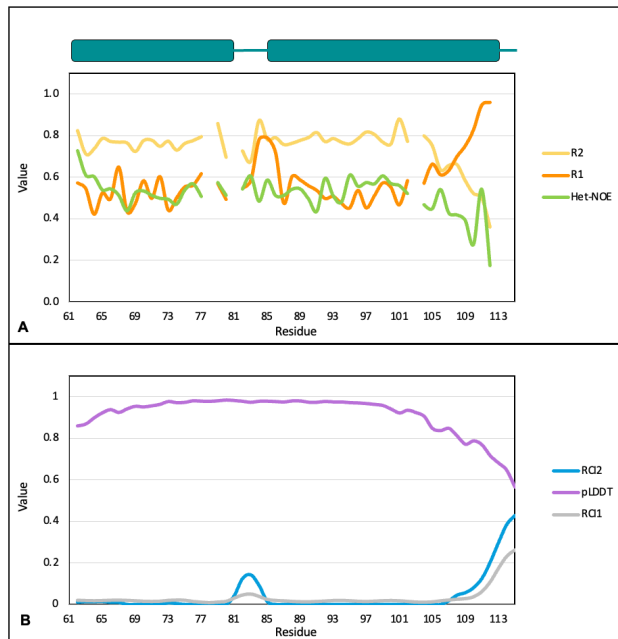
AF\_ALT\_3  
cluster 3



# Human DOC1 – Deleted in Oral Cancer



AlphaFold2-sample – models conformational dynamics that match well to experimental data



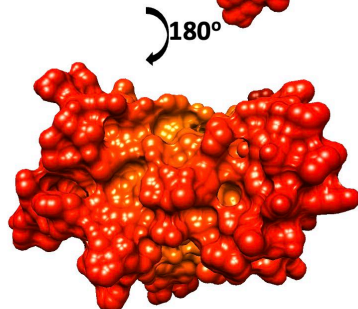
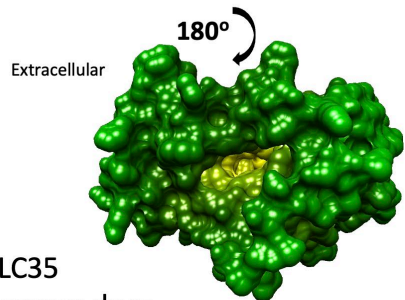
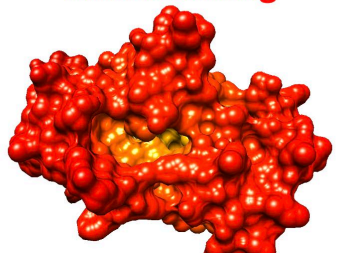
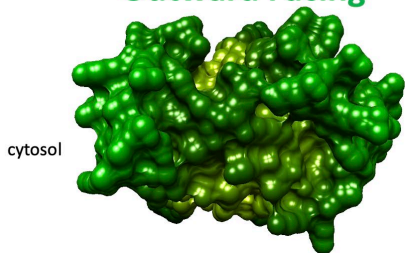
L Spaman  
YJ Huang  
R Tejero

# Modeling Alternative Conformational States of Pseudo-Symmetric Membrane Protein Transporters using Methods from Machine Learning

G.V.T. Swapna, Namita Dube, Monica J. Roth, and Gaetano T. Montelione

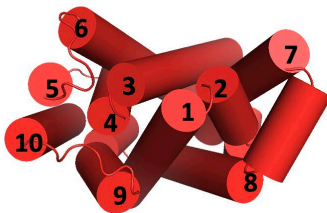
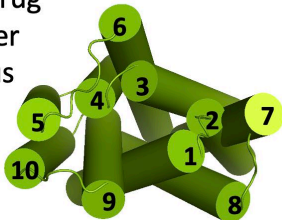
Outward Facing

Inward Facing

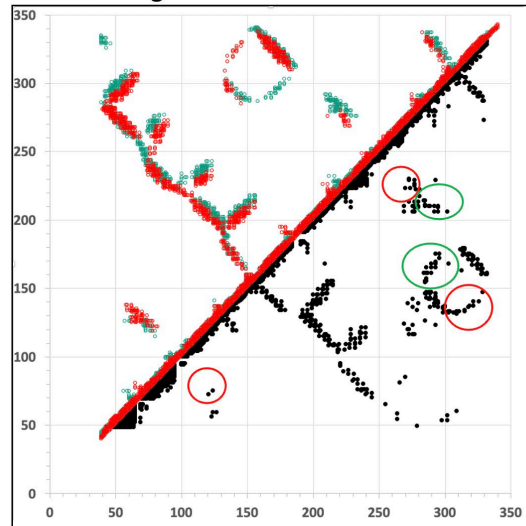


SLC35

- cancer drug transporter  
- MLV virus receptor



AF-alt predicts both outward and inward facing conformations



Some ECs are consistent only with outward facing – some only with inward facing

Mutations based on ECs can be used to shift preference from outward to inward



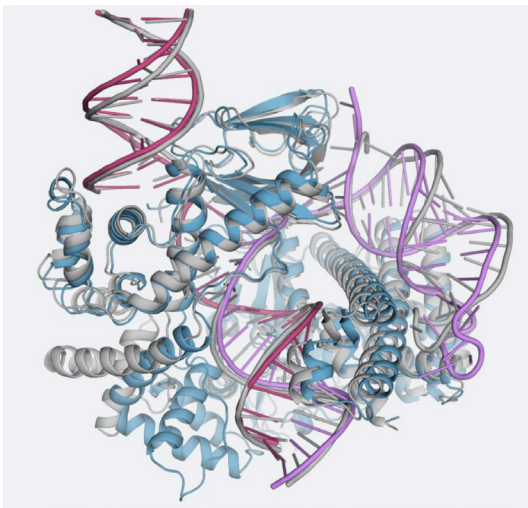
# Performance and structural coverage of the latest, in-development AlphaFold model

Google DeepMind AlphaFold Team<sup>1</sup> and Isomorphic Labs Team<sup>2</sup>

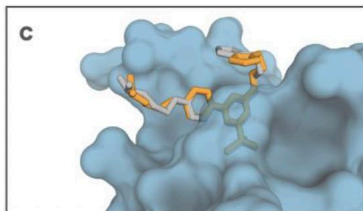
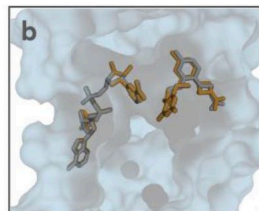
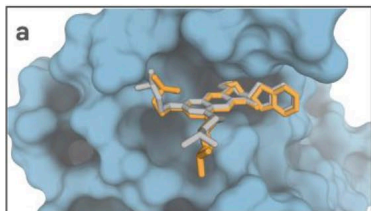
<sup>1</sup>DeepMind, London, UK, <sup>2</sup>Isomorphic Labs, London, UK

bioRxiv

Nov 2023



Protein:Nucleic Acid  
Complex: AF3 vs  
ground truth



Small molecule binding: AF3 vs ground truth



# BIG DATA

# RPI Structural Bioinformatics Lab

The Rensselaer Polytechnic Institute: Structural Bioinformatics Laboratory carries out structure and function studies of genomically-defined protein targets. The lab uses both NMR and X-ray crystallography, along with computational methods, to address questions emerging from genomic and functional genomic analyses. The lab also develops software to automate structure determination by NMR methods.

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YJ Huang

T Ramelot

GVT Swapna

R Tejero

G Liu

T Acton

B Shurina

L Spaman

N Dube

A DeFalco

L Klang

R Greene-Cramer

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